

# Successful extrapolation of paracetamol exposure from adults to infants after oral administration of a paediatric aqueous suspension is highly dependent on the study dosing conditions

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Running title:

Extrapolating drug exposure from adults to infants

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## 1 Abstract

2 Extending licensed drug use to the pediatric population has become an essential part of the drug  
3 development process. Nonetheless ethical concerns limit clinical testing in paediatric populations and  
4 data collected from oral bioavailability and food effect studies in adults are often extrapolated to the  
5 target paediatric (sub)populations. However, based on published information, food effects on drug  
6 absorption in infants may not be adequately evaluated by data collected in adults. In the present study,  
7 a physiologically based pharmacokinetic (PBPK) approach for modeling paracetamol suspension data  
8 collected in adults was proposed with the ultimate aim to investigate whether extrapolation to infants  
9 is substantially affected by the dosing conditions applied to adults. The development of the PBPK  
10 model for adults was performed using GastroPlus™ and, after scaling to infants considering  
11 physiological, anatomical, and drug clearance changes, extrapolation of the different dosing conditions  
12 was performed by applying dosing conditions dependent changes on the paracetamol gastric emptying  
13 process. Successful predictions of observed plasma concentration levels in infants were achieved when  
14 extrapolating from fasted and infant-formula-fed conditions data. Data collected following the  
15 reference meal appeared less useful for simulating paracetamol suspension performance in infants.  
16 The proposed methodology deserves further evaluation using high-quality clinical data both in adults  
17 and in infants.

## 18 Introduction

19

20 Extending licensed drug use to the paediatric populations has become an essential part of the drug  
21 development process to ensure appropriate dosing, efficacy and safety from birth to adulthood (1,2).  
22 As in adults, the oral route of administration is preferred from birth to adolescence and bioavailability  
23 studies are required to ensure suitable drug exposure and drug pharmacokinetics (PK) following the  
24 administration of the age-appropriate dosage form. However, ethical concerns and recruitment issues  
25 limit clinical testing in these vulnerable age groups more than in adults (3–5).

26

27 Physiologically based pharmacokinetic (PBPK) modelling could be a useful tool to drastically decrease  
28 the need for performing clinical studies in paediatric populations and, therefore, largely eliminate  
29 relevant concerns. Based on the ability to create PBPK physiologies representative of various human  
30 developmental stages, PBPK modeling utilization in paediatrics can facilitate drug performance  
31 predictions prior to testing in a clinical setting and guide drug formulation development (3,5).  
32 Additionally, to date, PBPK modeling has proven valuable as a tool to gain mechanistic understanding  
33 of physiological and drug parameters governing oral absorption processes across various paediatric  
34 age ranges (3,6–9). Interestingly, however, only a few of these studies use multi-compartmental  
35 representation of the gastro-intestinal (GI) tract, while implementing age-dependent physiological and  
36 anatomical changes to investigate different dosing and prandial conditions in the target population  
37 (3,8).

38

39 PBPK model development procedure to extrapolate adult data to paediatric populations employs a  
40 stepwise workflow, beginning by building a validated adult disposition model, followed by the  
41 development and validation of an adult absorption model, and, ultimately, the extrapolation to the  
42 paediatric population of interest (10). A recent draft guidance by the US Food and Drug Administration  
43 (FDA) proposed the use of age-specific meals and quantities for the investigation of populations  
44 receiving specific meals, e.g., infant formula for infants, without specification of an exact quantity (11).  
45 Although several studies in adults have employed infant formula or soft foods (e.g., applesauce,  
46 yoghurt, and fruit puree), the age-adjusted meal quantities simulate drug product administration with  
47 small amounts of food to facilitate drug formulation dosing and improve acceptability, rather than  
48 investigate the potential impact of dosing conditions on drug product performance (3,12). A recent  
49 study in healthy adults revealed reduced early exposure of paracetamol and ibuprofen, after  
50 administration under conditions simulating the fed state infants and toddlers (1-24 months) compared

51 to the administration under conditions simulating the fasted or fed state conditions in adults, as  
52 suggested by the current regulatory guidelines (9,11,13,14).

53

54 This manuscript describes and evaluates a PBPK modeling approach for extrapolation of drug exposure  
55 form adults to infants with view to the different conditions that can be used to inform the modeling  
56 process. The first objective was to propose a PBPK approach for modeling the recently collected  
57 paracetamol paediatric suspension data in adults under fasted and fed state conditions (13,14), and,  
58 under conditions mimicking dosing to infants (9). The second objective was to investigate if  
59 extrapolation to infants was substantially affected by the dosing conditions applied to adults. Both  
60 objectives were achieved by using the PBPK modeling platform GastroPlus™ V9.7.

## 61 Methods

### 62 PK data collection

63 Initially, a thorough search at PubMed was performed (completed March 2020) for previously  
64 published plasma data after intravenous administration (bolus and infusion) and oral administration  
65 (solution and suspension forms) of paracetamol to adults and paediatrics. Data that had been collected  
66 after administration of liquids containing excipients influencing the product performance, from an  
67 unspecified product, after co-administration with drug(s) influencing the GI physiology, and/or by  
68 employing paediatrics without age stratification were excluded from further consideration. The  
69 Statelova *et al.* (2020) study in adults was used as the basis for extrapolation to paediatrics. In addition  
70 to that study (9), a total of 23 paracetamol PK studies met the search criteria, with 15 studies in adults  
71 and 8 in paediatrics. From the adult studies, 12 studies reported i.v. paracetamol administration (15–  
72 24) and 2 studies reported oral administration of paracetamol solutions in the fasted state (19,21).  
73 From the 8 paediatric studies, 5 reported i.v. administration in infants (1 month – 2 years), children  
74 and adolescents (2 - 18 years) (25,26) and 2 studies reported oral administration in infants (1 month-  
75 2 years) or infants and young children (3-36 months) (27,28). From the 23 studies retrieved from  
76 literature, plasma concentration-time profiles and respective standard deviations (SD) or standard  
77 errors of the means (SEM) were digitized using the WebPlotDigitizer software V4.1 (Ankit Rohatgi,  
78 2017). Along with the reported plasma levels as a function of time, extracted information also included  
79 drug dosing conditions, drug products, and demographics of the study population, i.e., number of  
80 study subjects, age, gender, body height, body weight, and race. For the Statelova *et al.* study (2020)  
81 (9), in addition to the published mean plasma concentrations and demographics, individual data were  
82 also available.

83

### 84 Modeling strategy

85 The PBPK model for paracetamol was developed using the GastroPlus™ software platform (V. 9.7,  
86 Simulations Plus, Lancaster, CA, USA). The model development strategy employed a “middle-out”  
87 approach (29), whereby model parameterization was guided by clinical observations in humans  
88 (Figure 1). As part of the applied “learn-confirm-apply” approach (30), the model was built and refined  
89 using *in vivo* data sets and, then it was verified using external data sets before applying/extrapolating  
90 to infants. As a first step, a disposition model for healthy adults was developed and optimized  
91 according to clinical studies after i.v. drug administration reported in literature (16), followed by  
92 verification with external clinical datasets not used for the model development (15,18). After gaining  
93 certainty in the disposition model, oral absorption in adults was described using the Advanced

94 Compartmental Absorption and Transit (ACAT™) model within the GastroPlus™ platform for liquid drug  
95 formulations i.e., solution and suspension. For the paracetamol suspension formulation, different  
96 prandial and dosing conditions were modeled and relevant parameters were adjusted according to  
97 data observed in adults (9). The model was scaled to different paediatric age groups for which clinical  
98 data following intravenous drug dosing was available to confirm the scaling of drug disposition across  
99 ages. Finally, different dosing and prandial conditions for the administration of the paediatric  
100 suspension were extrapolated from adults to infants and compared to data observed in this paediatric  
101 population.

102

### 103 Adult model

104 A full PBPK model for adults was established for paracetamol using the data listed in Table I. Human  
105 physiologies matching to each simulated study demographics (age, body weight, gender, body-mass-  
106 index) were created using the Population Estimates for Age-Related (PEAR) Physiology module within  
107 GastroPlus™ (6,43,44). Within the PEAR™ physiology module, after selecting the subject  
108 demographics, blood flows, organ and tissue sizes, as well as tissue composition are adjusted based on  
109 literature (6,43,44). A default physiology for a healthy American adult 30-year-old male with a body  
110 weight of 70 kg was used when the simulated study lacked reporting of the demographics. A study  
111 reported by Clements *et al.* investigated the i.v. administration of paracetamol at 5 mg/kg and 20  
112 mg/kg doses covering the range of the typical paracetamol dose-strengths, e.g. 15 mg/kg (16).  
113 Additionally, the study has been used successfully for building paracetamol PBPK models in literature  
114 and the study report allowed for reliable extraction of the datapoints (32,33,45). Therefore, based on  
115 data sets from the study by Clements *et al.* (16), clearance (CL) and volume of distribution at steady  
116 state (V<sub>ss</sub>) were estimated via non-compartmental analysis performed with the PKPlus™ module  
117 within GastroPlus™ and were used as benchmark values for CL and V<sub>ss</sub> in healthy adults. Within the  
118 current modeling development, V<sub>ss</sub> was derived from the tissue partitioning coefficient values (K<sub>p</sub>) for  
119 perfusion-limited tissues estimated using the Rogers, Roland, Lukacova method (6,38). The predicted  
120 V<sub>ss</sub> value was adjusted to match the benchmark value from clinical observations (Table I). The *in vivo*  
121 clearance was scaled to *in vitro* clearance for each enzyme contributing to drug metabolism using a  
122 retrograde stepwise routine (46) as briefly explained in the following text (exact calculations are  
123 provided as Supplementary Information). Based on the extensive liver metabolism of the drug and the  
124 literature reports indicating insignificant paracetamol metabolism in the gut and kidney (16,22,47,48),  
125 the total clearance was considered to originate from the liver. Hence, the benchmark total  
126 paracetamol clearance after i.v. administration was used for the estimation of the *in vivo* unbound  
127 intrinsic hepatic clearance according to the well-stirred clearance model (49). Based on the hepatic

128 metabolism contributions of isoenzymes of the Cytochrome P-450 (CYP), UDP-glucuronosyltransferase  
129 (UGT), and cytosolic sulfotransferases (SULT) enzyme families, *in vivo* intrinsic clearance values per  
130 isoenzyme were calculated (7,16,32). These were further employed to determine *in vitro* drug-  
131 metabolizing enzyme parameters (Table I) (7,32,33). The disposition model was verified with data from  
132 reported i.v. studies of paracetamol (External Datasets) that were not utilized for the model  
133 development.

134  
135 The ACAT™ model describes the drug dissolution, precipitation, and luminal absorption during drug  
136 transfer through the nine compartments of the GI-tract within the model, i.e. stomach, duodenum,  
137 two jejunum, three ileum, and colon compartments (50,51). Each compartment is characterized by a  
138 physiology-adjusted small-intestinal (SI) length, radius, specific absorption factor (ASF), intraluminal  
139 fluid volumes and composition, and transit times. Human effective permeability of paracetamol  
140 ( $P_{eff,man}$ ) was estimated from the *in vitro* apparent permeability in Caco-2 cells ( $P_{app,Caco2}$ ) employing  
141 atenolol as a calibrator (7,34,35), Eq. 1.

$$\log P_{eff,man} = 0.6795 \times \log P_{app,Caco2} - 0.3036 \quad \text{Eq. 1}$$

142  
143  
144  
145 Oral solution data from literature were used as confirmation that the estimated permeability predicted  
146 paracetamol oral absorption (19,21). The software's default gastric transit time (GTT) value of 0.1 h  
147 and 1<sup>st</sup> order gastric emptying (GE) kinetics were employed for the solution; GTT for 1<sup>st</sup> order emptying  
148 kinetics represents the mean gastric transit time value (MGTT) defined as the GE half-life divided by  
149 the natural logarithm of 2.

#### 151 Modeling under different dosing conditions

152 The exploratory relative bioavailability study by Stelova *et al.* was performed in healthy adult male  
153 volunteers and included three study arms to investigate suitable dosing conditions to evaluate the  
154 performance of paediatric suspensions for administration in infants (1 month-2 years), i.e.,  
155 paracetamol paediatric suspension (Panadol®) (9). The human physiology used for the modeling  
156 represented the average values of 78 kg, 28 years of age and BMI of 20.23 kg/m<sup>2</sup> as reported in the  
157 study by Stelova *et al.* (mentioned throughout the text as "population representative"). A single dose  
158 of 1000 mg was administered on a crossover basis under different dosing conditions. In particular, the  
159 investigated dosing conditions included administration of the paediatric drug formulation under fasted  
160 conditions, fed conditions as proposed by current regulatory guidelines for adults (30 min after the  
161 start of the consumption of the reference meal) (13,14) and conditions mimicking dosing in infants

162 where the drug formulation was administered during infant formula consumption, i.e. infant-formula-  
163 fed conditions (9).

164

165 Model parameters were adjusted to capture the performance of the paediatric formulation as  
166 observed in adults, e.g., adjustment of GTT as GE and arrival of paracetamol in the SI were associated  
167 to paracetamol appearance in the systemic circulation (52). Due to the multiple-peak phenomenon  
168 observed for Panadol® under fasted conditions, an empirical modeling strategy was employed  
169 following “mixed multiple dosing” (MMD) of the suspension to verify that gastric emptying events were  
170 responsible for the observed profile shape (and not other absorption factors). Multiple GI-physiologies  
171 were created and applied using alternating rapid (GTT 0.1 h or 0.25 h) and slow GTT (10 h) values  
172 starting at different timepoints after drugs administration within the performed simulation; the  
173 multiple GI-physiologies and the different GTT introduced were adjusted (fitted) to simulate the  
174 observed discontinuous GE of the suspension under fasted conditions. As the goal was to extrapolate  
175 the model to infants, a compromise was made for a single GE process for fasted state modeling.

176

177 For simulations of paracetamol dosing under postprandial conditions, the hepatic blood flow was  
178 increased by one third of the baseline hepatic blood flow, to mimic the increase splanchnic blood flow  
179 observed after food consumption (50). By switching the prandial conditions option to “fed conditions”,  
180 the luminal conditions within the simulated adult physiology were adjusted to the fed state e.g., bile  
181 salt increased as a function of fat content in the meal, pH increase in the gastric compartment, and  
182 prolongation of GE. To simulate different prandial conditions within GastroPlus™ V 9.7, in addition to  
183 a single default fed options for fed conditions applied in previous software versions, the “user-defined  
184 meal” option allows for flexibility in adapting the GTT as a function of the caloric content and the bile  
185 salt level adjustment according to the percentage of fat of the selected meal. Simulations were  
186 performed according to the software-proposed values for the different prandial conditions (referred  
187 to as “default settings or conditions” simulation throughout the manuscript). Under fed conditions,  
188 the total caloric content of the meal was 990 kcal with 60 % derived from fats, while under infant-  
189 formula-fed conditions the total caloric content was 520 kcal with 43 % fat content (9). Within the  
190 present model development, adjustments were undertaken based on *in vivo* observations for  
191 parameters that changed as a function of the meal texture and formulation type, e.g., following the  
192 solid-liquid reference meal the GE process followed 1<sup>st</sup> order kinetics, although incomplete mixing of  
193 the suspension led to shorter paracetamol GTT compared to typically reported GE times for similar  
194 meals, or GE times were prolonged and GE followed zero order kinetics when administered with the  
195 infant formula (liquid homogeneous) (9). It should be noted that under zero order GE kinetics the GTT  
196 value to be entered in the software represents the time for drug gastric emptying to complete.

197

198 Moreover, population simulations were performed for the refined settings for the three dosing  
199 conditions using a virtual population with similar demographics to the study by Stelova *et al.* (9).  
200 Under each type of dosing conditions, eight virtual male healthy subjects were generated using the  
201 PEAR™ module of the modeling platform with age range 21-48 years, body weight range 60-100 kg,  
202 and BMI range 20-28 kg/m<sup>2</sup>. Simulation were performed over 10 h. Software default variability was  
203 employed for all parameters (44), except for the GTT values employed under both postprandial  
204 conditions, for which no variability could be included based on software limitations.

205

### 206 Paediatric model scaling

207 Paediatric physiologies were generated for each paediatric study using the PEAR physiology module  
208 within the modeling platform (44), i.e., a mean population representative according to the study  
209 demographics (25,27,28). The generation of virtual subjects (using the PEAR physiology module)  
210 accounted for maturation and development changes occurring from birth to adolescence, i.e., body  
211 and tissue sizes, blood parameters, tissue compositions, as well as hepatic CYP-enzyme abundances  
212 based on data from an exhaustive literature review (6,43,44,53). The microsomal protein per gram  
213 liver tissue was assumed to be independent of age (44), while age-adjusted scaling factors for enzymes  
214 of the UGT and SULT families were extracted from literature to scale the adult baseline abundances  
215 incorporated in the systemic adult parameters within the simulation platform (32,33,53,54). The  
216 presented scaling approach has been shown to lead to successful modeling of paracetamol metabolism  
217 across different ages following intravenous drug administration (33). Clinical observations in infants  
218 and in children and adolescents after i.v. administration were used to verify the present disposition  
219 model in paediatrics (25).

220

### 221 Oral absorption in infants under different dosing conditions

222 Relevant to oral absorption modeling as a function of age, the change of PEAR physiologies accounts  
223 for developmental changes in the paediatric GI tract within the ACAT™ physiology, such as GI-segments  
224 length and transit times, and accounts for some of the age-dependent factors that can influence  
225 paracetamol bioavailability. For the extrapolation to infants and evaluation of the usefulness of the  
226 three dosing conditions applied in the study by Stelova *et al.*, adjusted parameters from the adult  
227 paediatric suspension model were scaled to infants and applied to paediatric simulations. In the  
228 dataset described in (27), 5 infants with a mean age of four months (2 - 6 months) were dosed with a  
229 target dose of 15 mg/kg Calpol® suspension (dose administered 19.6 mg/kg), while in the dataset  
230 reported in (28) the paracetamol dose 12.14 mg/kg was given to infants and young children with a

231 mean age of ten months (range: 3 - 36 months). As in adults, the performance of the software default  
 232 settings was evaluated during the infant model development, i.e., default settings for the fasted state  
 233 and “user-defined meal” settings using zero and first order kinetics, for a solid-liquid meal and liquid  
 234 homogeneous meal, respectively. As a next step, extrapolated parameters based on the refined adult  
 235 model according to the study by Statelova *et al.* were used as input for the simulations in infants, with  
 236 detailed description of the extrapolation rationale for the three different conditions described in the  
 237 following paragraph (9).

238

239 A recent meta-analysis of GE as a function of age revealed that food type rather than age determined  
 240 GE across ages (55). Therefore, under the assumption that no age dependent GTT changes would occur  
 241 under fasted conditions, the GTT value found to appropriately describe the fasted state was inherited  
 242 directly from the refined adult PBPK model. For the fed conditions and the infant-formula-fed  
 243 conditions, the average paracetamol meal-dependent GE rate was estimated as a function of the type  
 244 (solid-liquid vs. liquid homogeneous meal) and the caloric content of the meal. For this purpose, the  
 245 caloric content of the meal given to adults was divided by the adjusted GTT values employed for the  
 246 fed and infant-formula-fed conditions found to best describe paracetamol appearance in the systemic  
 247 circulation (Eq. 2). Subsequently, fed GTT values for infants were estimated based on the caloric  
 248 content of the recommended formula amounts for the age of interest and the paracetamol meal-  
 249 dependent GE (Eq. 3). Different recommended meal calories reported for infant formula were selected  
 250 for the infant group with a mean age four months and for the infant/young children group with a mean  
 251 age ten months, i.e., 140 kcal and 170 kcal, respectively (3).

252

$$253 \quad \text{Average Paracetamol Gastric emptying rate}_{adults,Meal} = \frac{\text{Caloric content (meal based)}}{\text{Paracetamol GTT(meal based)}} \quad \text{Eq. 2}$$

254

$$255 \quad \text{Paracetamol GTT}_{infants,meal} = \frac{\text{Meal caloric content recommended for age needs}}{\text{Average Paracetamol Gastric emptying rate}_{adult,Meal}} \quad \text{Eq. 3}$$

256 where Paracetamol GTT represents the MGTT for a first order GE process (solid-liquid meal) and total  
 257 GTT for a zero order GE process (Infant formula).

258

259 In addition to the single simulations, population simulations were performed for the two infant study  
 260 groups, matching the demographics from each study (27,28) under the three dosing conditions  
 261 employing the adjusted GTT values. Software limitations to parameter variability incorporation (GTT)  
 262 is as described for the adult population simulations.

263

## 264 Model performance evaluation

265 For population representative and population simulations, (mean) predicted and observed PK  
266 parameters describing total drug exposure, peak exposure, and time to reach peak exposure (area  
267 under the plasma concentration vs. time curve (AUC), C<sub>max</sub>, and T<sub>max</sub>, respectively) were compared  
268 using the predicted vs. observed fold difference ( $FD_{pred/obs}$ ). The predicted concentration-time profiles  
269 from population representative simulations and mean predicted plasma concentration-time profiles  
270 from the population simulations were evaluated by the average fold error ( $AFE$ ) and the absolute  
271 average fold error ( $AAFE$ ) calculated using Equation 4 (Eq. 4) and Equation 5 (Eq. 5), respectively.

272

$$273 \quad AFE = 10^{\left(\frac{1}{n} \sum \log \left(\frac{PRED_i}{OBS_i}\right)\right)} \quad \text{Eq. 4}$$

$$274 \quad AAFE = 10^{\left(\frac{1}{n} \sum \left| \log \left(\frac{PRED_i}{OBS_i}\right) \right| \right)} \quad \text{Eq. 5}$$

275

276 where n denotes the number of observed sampling points, PRED<sub>i</sub> and OBS<sub>i</sub> denote the predicted and  
277 observed plasma concentration, respectively, at the sampling time point i.

278

279 Additionally, for the population simulations, 90 % confidence intervals (CI), and probability contours  
280 (10 %, 25%, 50 %, 75 %, 90 %, 95 % and 100 %) including 5<sup>th</sup> and 95<sup>th</sup> percentiles were evaluated.

281

282  $AFE$  values indicate the trend of the simulated data to underpredict ( $AFE < 1$ ) or overpredict ( $AFE > 1$ )  
283 the observed plasma concentrations, while an  $AAFE$  value close to unity signifies the precision of the  
284 simulations. Predictions resulting in  $FD_{pred/obs}$  and  $AAFE$  values less than two are considered adequate  
285 (56), while stricter evaluation criteria for  $FD_{pred/obs}$  between 0.5-1.5 for and  $AAFE$  below 1.5 indicate  
286 a successful prediction (57).

287

## 288 Parameter sensitivity analysis

289 Parameter sensitivity analysis (PSA) was performed according to a one-factor-at-a-time approach to  
290 understand the uncertainties of the parameters employed within the refined adult oral absorption  
291 model developed and evaluated in the present investigation regarding drug-related parameters, i.e.,  
292 drug solubility, permeability, particle size radius, as well as physiological parameters, i.e., GTT. The  
293 investigation was performed with a population representative matching the mean demographic  
294 parameters of the clinical study by Stelova *et al.*, i.e. 28-year-old male with a 78 kg bodyweight (9).  
295 Additionally, PSA was run for physiological, drug-dependent, and dosing parameters contributing to  
296 model uncertainty for infants under fasted, fed, and infant-formula-fed conditions using a physiology

297 matching the mean infant representative in Hopkins *et al.* (27). Physiological parameters included SI  
298 radius and length, GTT, SITT, and gastric and duodenal pH, while drug-dependent parameters as  
299 permeability, bile salt solubilization ratio, diffusion coefficient, reference solubility, and particle size  
300 were investigated as drug-dependent parameters. Finally, the influence of dose strength and dosing  
301 volume were simulated to explore the influence of trial conditions. Under fed and infant-formula fed  
302 conditions, PSA was performed additionally regarding the caloric content of the paediatric meal  
303 administered to the infants (Table SI, Supplementary Information). The extent to which paracetamol  
304 PK and PK parameters are influenced by the selected parameter range was evaluated.

## 305 Results

### 306 Adult model performance

307 The developed disposition model for adults was able to adequately describe paracetamol disposition  
308 in the i.v. study used for modeling development when paracetamol was administered at a low dose  
309 5 mg/kg, i.e., 350 mg (*AAFE* 1.045) and high dose of 20 mg/kg, i.e., 1400 mg (*AAFE* 1.080) (Figure 2A  
310 and 2B, respectively). External datasets used for model verification from two studies (15) simulated  
311 the observed data acceptably (*AAFE* 1.131) for predictions at low paracetamol dose of 500 mg  
312 paracetamol and for predictions at high paracetamol dose of 1000 mg paracetamol (*AAFE* 1.212), as  
313 shown in Figures 2C and 2D, respectively. Predicted clearance and  $V_{ss}$  values were within observed  
314 ranges reported in the literature (Table SII, Supplementary Information). In addition, the disposition  
315 model was found to simulate all clinical study data following i.v. administration reported in literature  
316 with reasonable accuracy, as shown in Figure S1 and Table SIII in the Supplementary Information. The  
317 effective permeability value for humans scaled from Caco-2 apparent permeability experiments was  
318 in line with reported permeability ranges (45,58,59). Using the default GastroPlus™ settings for oral  
319 solution including a GTT of 0.1 h, the developed model achieved satisfactory prediction of paracetamol  
320 exposure after oral administration of 1000 mg solution in healthy adults in two different clinical studies  
321 [*AAFE* 1.088, Figure 3A (19) and *AAFE* 1.275, Figure 3B (21)]; thus confirming the suitability of the  
322 permeability value applied (Table SIII, Supplementary Information).

323

### 324 Modeling under different dosing conditions

325 The default settings for fasted and fed conditions utilizing the user-defined meal option for defining  
326 the meal specific caloric (reference meal 990 kcal and infant formula 520 kcal) and lipid (reference  
327 meal 60 % and infant formula 43 %) content failed to describe the data observed for the paracetamol  
328 suspension administered to healthy adults (Figure 4A, 4C, 4E). Consequently, adjustments of the GTT  
329 values for fasted, fed, and infant-formula-fed conditions were undertaken to match data observed *in*  
330 *vivo* (Figure 4B, 4D, 4F). Results herein are presented for the population representative from the  
331 Stelova et al. clinical study (9), while results for population simulations including mean profiles and  
332 their respective 90 % CIs, 5<sup>th</sup> and 95<sup>th</sup> percentiles and probability contours are reported in the  
333 Supplementary Information in Figure S3 and S3-1 and the mean predicted PK parameters and their  
334 respective  $FD_{pred/obs}$  values are presented in Table SIV. Due to the multiple peak phenomena observed  
335 under fasted conditions in adults, drug performance was better described when multiple GE events  
336 were fitted using the MMD dosing available in the software (*AFE* 0.941 / *AAFE* 1.052, Figure S2).  
337 However, for the purposes of extrapolation to infants, a compromise was made for a single GE process

338 for fasted state modeling, employing an adjusted GTT of 0.75 h (*AAFE*1.200). In the fed state following  
339 the reference meal, the suspension was emptied faster than the proposed GE times for the reference  
340 meal, thus requiring an adjustment of the 3.43 h GTT proposed for the meal to 1.5 h. Simulations  
341 utilizing the adjusted GGT value indicated better predictions compared to predictions using default  
342 values for GTT, i.e., *AAFE*1.145 vs *AAFE*1.733, respectively. In line with typical GE kinetics of liquid  
343 meals (60), under infant-formula-fed conditions, mean plasma concentration-time profiles were well-  
344 described by a zero-order GE. As for the reference meal, GTT adjustments were needed, as default  
345 parameters underpredicted the delay observed with (*AAFE*1.059 vs *AAFE*1.873). For the population  
346 simulations, although the mean predictions successfully matched the observed data, individual  
347 measured plasma concentrations fell outside the 5<sup>th</sup> and 95<sup>th</sup> percentiles for the simulations (Figure  
348 S3, Supplementary Information). This was especially noticeable at early times (Figure S3A and C,  
349 Supplementary Information) for both fed conditions (reference meal and infant formula) and was  
350 attributed to the limitation of the platform to include any variability for the adjusted GTT values.

351

## 352 [Scaling to paediatrics](#)

### 353 [Disposition](#)

354 Disposition kinetics and clearance employing isoenzymes of the CYP, UGT, and SULT families could  
355 predict observed paracetamol levels following i.v. administration over 0.25 h at doses of 12.5 mg/kg  
356 or 19.6 mg/kg (7,32,33). The model scaling was suitable to predict reported concentrations for i.v.  
357 administration for a population representative of infants (male, mean age 4 months and 4 kg) and of  
358 a population representative of a mixed children and adolescents group (male, mean age 6 years old  
359 and 23 kg) (25). Simulations for population representative of infants were performed for a higher dose  
360 administered at 15 mg/kg (*AAFE*1.312, Figure 5A) and a lower dose administered at 12.5 mg/kg (*AAFE*  
361 1.081, Figure 5B). On the other hand, simulations for a population representative of the mixed group  
362 were adequate for a high dose of 15 mg/kg paracetamol (*AAFE*1.420, Figure 5C) and a lower dose of  
363 12.5 mg/kg paracetamol (*AAFE*1.187, Figure 5D).

364

### 365 [Oral absorption in infants](#)

366 Clinical data in infants following oral administration of a liquid paracetamol formulation available from  
367 two datasets were used for the evaluation of the usefulness of the developed adult model to predict  
368 paracetamol exposure in infants (27,28). Initially, using the default software settings, simulation of  
369 paracetamol plasma profiles in infants were performed under the three different dosing conditions.  
370 Then, for the purpose of extrapolating the fed conditions and the infant-formula-fed conditions to

371 infants, adjusted GTT values for infants were calculated based on these values and on caloric needs of  
372 the population representative of each study (Eq. 2 and Eq. 3), presented in Table II.

373  
374 All simulations performed for the infant population representative, i.e. 4 month-old infants according  
375 to (27), are presented in Figure 6. Fasted state simulations employing default software parameters  
376 (GTT 0.1 h) could not describe early drug exposure, as they underpredicted Tmax ( $FD_{pred/obs} = 0.60$ ) and  
377 overpredicted Cmax ( $FD_{pred/obs} = 1.3$ ), although the overall description of the postabsorptive phase was  
378 adequate ( $AAFE 1.185$ ). The fasted conditions extrapolated from the refined adult model (GTT 0.75 h)  
379 led to a better prediction of the Tmax and slight underprediction of Cmax ( $FD_{pred/obs} = 0.90$ ), capturing  
380 both the early and the overall exposure better than the default settings (Figure 6A vs 6B, Table III).  
381 Following first order GE kinetics of the reference meal (a solid-liquid meal) and a caloric content of  
382 140 kcal (the caloric content of a meal for a 4-month-old infant), default simulations (GGT = 2.1 h)  
383 could not successfully describe the data observed ( $AAFE 1.523$ , Figure 5C). Calculation of the adjusted  
384 GTT for infants resulted in a value of 0.21 h (Table II) and although the postabsorptive PK were captured  
385 ( $AAFE 1.201$ ) the early exposure was overpredicted (Figure 5D). Under infant-formula-fed conditions  
386 and following zero order GE kinetics (as in adults), default simulations (GGT = 2.1 h) inaccurately  
387 described the data observed ( $AAFE 1.428$ , Figure 5C). However, when using the adjusted GTT value  
388 (1.21 h), successful predictions of both early exposure and total exposure were achieved ( $AAFE 1.215$ ,  
389 Figure 6F). Mean simulation profiles from the population simulations (n=25, age range 2-6 months)  
390 corroborated the observations from the single simulations, as shown in Figure S5 (Supplementary  
391 Information).

392  
393 Similarly to the simulations for younger infants (27), early exposure was overpredicted when applying  
394 software default parameters for the fasted state in infants with mean age of 10 months (28) and  
395 resulted in inaccurate predictions ( $AAFE 1.442$ , Figure 7). In contrast, fasted conditions using the  
396 refined adult model (GTT 0.75 h) matched observed data well ( $AAFE 1.201$ , Figure 7B). Following first  
397 order GE kinetics of the reference meal and caloric content of 170 kcal (the caloric needs of a 10-  
398 month-old infant), simulations employing default value for GTT = 1.89 h resulted in greater absorption  
399 delay than observed *in vivo*, as indicated by the  $AAFE$  value of 1.87 (Figure 7C). The use of the adjusted  
400 GTT value for this study (Table II), although seeming to better predict the overall oral paracetamol  
401 performance compared to the default GTT values ( $AAFE 1.274$  vs  $AAFE 1.87$ ) led to overprediction of  
402 Cmax ( $FD_{pred/obs} = 1.59$ ). Under infant-formula-fed conditions, following zero order GE kinetics, default  
403 software settings (GTT 1.89 h) and adjusted GTT (1.47 h) underpredicted early exposure, however,  
404 employment of the adjusted GTT value showed slight improvement in the overall prediction compared  
405 to the default settings ( $AAFE 1.40$  vs  $AAFE 1.695$ , Figure 7F and 7E, respectively). Population

406 simulations performed in 25 virtual subjects aged 3-36 months (28) indicated similar findings as the  
407 observations based on the single simulations with the mean population representative (Figure S7,  
408 Supplementary Information).

409

#### 410 [Parameter sensitivity analysis](#)

411 PSA was performed for permeability and GTT under the three dosing conditions for the refined model  
412 for an adult population representative and a 4-month-old infant (9,27). Paracetamol PK showed  
413 sensitivity regarding the effective human permeability both in infant and adult population  
414 representatives, especially under fasted conditions (Figures S8 and S9, Supplementary Information).  
415 Decrease in paracetamol permeability negatively influenced the fraction of drug absorbed with up to  
416 10 % compared to the baseline values (data not shown). Increase of GTT in adults and in infants  
417 resulted in lowered early exposure (Figure 8A), with prolonged Tmax values and Cmax decrease  
418 (Figure 9 and Figure S10, Supplementary Information). Furthermore, in infants, increased caloric  
419 content of the food translated into greater GTT values and led to more pronounced delay in  
420 paracetamol absorption under adjusted infant-formula-fed conditions when compared to  
421 extrapolation under adjusted fed conditions (Figure 8B and C and Figure 9 B and D). Overall,  
422 permeability and GTT changes demonstrated minor impact regarding total drug exposure.  
423 Additionally, reference solubility, bile salt solubilization ratio, dose volume, as well as the physiological  
424 parameters investigated demonstrated minor to no sensitivity in infants regardless of the dosing  
425 conditions applied, i.e. fasted, fed, or infant-formula-fed conditions (Table SI, Supplementary  
426 Information).

## 427 Discussion

428 Although PBPK modelling has been commonly used for the extrapolation from adults to paediatric  
429 populations, the usefulness of incorporating adult and/or infant-meal food effect data into PBPK  
430 modeling to extrapolate to infants has to the best of our knowledge not been reported yet. In this  
431 study, extrapolation to the infant paediatric subpopulation was performed based on the results of an  
432 exploratory clinical investigation of the paediatric paracetamol suspension in adults, which was  
433 designed to elucidate the effects of three different dosing conditions on drug performance, i.e., fasted,  
434 reference meal-fed and infant-formula-fed conditions (9). The applied PBPK modeling approach  
435 involved initial refinement of the adult oral absorption model for the different dosing conditions to  
436 match the *in vivo* observations reported by the Stelova *et al.* and these conditions were subsequently  
437 scaled to simulate paracetamol plasma concentration levels in infants observed after oral  
438 administration of paracetamol liquid formulations (27,28).

439  
440 The discrepancy between predictions using default software values and predictions following  
441 adjustment of GTT values based on observed product performance highlighted the importance of  
442 model refinement that considered *in vivo* data collected under age-relevant dosing conditions using  
443 the commercially available paediatric formulation (Figure 4). Although PBPK modeling confidence with  
444 respect to oral drug absorption in adults has increased over the years and is considered to be reaching  
445 maturation for children (3,7,8), some aspects of GE and SI-transfer might not be accurately captured  
446 by a default approach regardless of age, i.e., discontinuous GE of liquid formulations and/or mixing  
447 processes between drug formulation and meal (61). In particular, the mismatch between the fasted  
448 state default prediction and observations for the suspension in adults could be explained by  
449 discontinuous GE-events resulting in a prolonged GE of the suspension as opposed to a single rapid GE  
450 event assumed for liquid formulations (9,19,62), i.e., Figure 2. The software platform enabled modeling  
451 of GE times for the administered drug as a function of different meal caloric contents, assuming  
452 homogeneous mixing of the drug and ingested meal. However, the default software assumptions of  
453 homogeneous mixing between drug formulation and the administered meals did not adequately  
454 reflect paracetamol GE patterns (63,64). Incomplete mixing of the formulation with the solid-liquid  
455 reference meal would lead to faster paracetamol emptying compared to the meal, as observed in the  
456 simulations (Figure 4C and 4D). On the other hand, paracetamol suspension mixes better with the  
457 liquid homogeneous infant formula, leading to paracetamol GE predominantly together with the infant  
458 formula bolus (9,63). It should be noted that, under both postprandial dosing conditions,  
459 independently of the meal texture, distinct paracetamol GE processes were not accurately reflected  
460 by the default ACAT™ model (9,63).

461

462 The present infant paracetamol PBPK model was discussed with focus on absorption parameters, as  
463 the paracetamol disposition and clearance parameters across ages employed in the model have been  
464 verified and discussed elaborately in previous works (7,32,33). In the present study, successful  
465 predictions were achieved for 4 month old infants (27) utilizing the refined model based on the *in vivo*  
466 performance of paracetamol suspension in adults under fasted conditions or infant-formula-fed  
467 conditions based on the recommended age-adjusted meal caloric content for the calculation of GTT in  
468 infants (Eq. 2 and Eq. 3), as shown in Figure 6B and 6F and summarized in Table III. Simulation of the  
469 administered dose in the population representative of the second available study [mean age of  
470 10 months, (28)] led to most reasonable predictions using the refined model parameters for fasted  
471 conditions adjusted according to the study by Statelova *et al.* (Figure 7B and Table III). Similar  
472 observations resulted from population simulations for the adjusted dosing conditions (Figures S5 and  
473 S7, Supplementary Information), despite the simulation limitations based on the lack of variability  
474 included for GTT under both fed conditions. Although the prandial state in both infant studies was not  
475 reported, the adequacy of the predictions assuming infant-formula-fed state in a 4-month-old infant  
476 representative can be corroborated by the frequent feedings resulting in non-fasted conditions  
477 observed in young infants when compared to children and older age-ranges (3,65,66). In comparison,  
478 another age-dependent oral absorption modeling exercise employing default values for fasted and fed  
479 state in infants assuming a liquid feed and a semi-solid feed predicted slower absorption compared to  
480 the predictions in the present investigation (7,27). The delay in predicted absorption might be  
481 explained by the lack of meal size adjustment as a function of age and/or imperfect capturing of mixing  
482 events between formulation and meal. Lastly, within the current investigation, the extrapolation based  
483 on paracetamol GE kinetics after the ingestion of reference meal in adults (9) and the recommended  
484 age-adjusted meal calories for the estimation of GTT in infants resulted in overprediction of early  
485 exposure and rapid paracetamol absorption unlike the data observed in infants (27,28), thus appearing  
486 less suitable for the prediction of oral drug performance in infants.

487

488 In adults the usefulness of paracetamol as a GE marker to elucidate physiological events has been  
489 widely recognized under fasted state conditions (52,67), however, not after the high-calorie, high-fat  
490 meal recommended by regulatory agencies for the fed state (52,67). Within the present investigation  
491 of the fasted state in infants, when comparing the adjusted GTT value extrapolated from adults in the  
492 fasted state (GTT 0.75 h), the presence of thickening excipients in the paracetamol paediatric  
493 suspension could be the cause of delayed GE compared to GE of water in paediatrics, as in adults. As  
494 a note, reported GTT values in neonates who received 5 mL/kg non-caloric liquid and in infants who

495 received 20 mL/kg distilled water have been reported to be shorter, 0.17 h and 0.36 h, respectively,  
496 (68,69), but a meta analysis across paediatric ages determined a GTT of 0.75 h for aqueous solutions  
497 in the fasted state (55,70). Regarding the infant-formula-fed conditions, the adjusted paracetamol GTT  
498 values cannot be compared with reported values from physiological studies in infants following infant  
499 formula/milk feeds, because the GE kinetics in those studies are not always reported and/or different  
500 infant formula types, caloric amounts, and formula compositions are used (3,71–74).

501

502 As PBPK modeling scaling of oral absorption processes to paediatrics relies on several assumptions  
503 originating in knowledge gaps regarding physiological development and maturation in paediatrics (6),  
504 parameters crucial for oral absorption and their impact on drug exposure in infants were investigated  
505 using a one-factor-at-a-time PSA approach with primary focus on the prandial conditions. Drug  
506 (formulation) related parameters and most physiological changes in infants appeared to be less  
507 important for paracetamol exposure (Table SI) (45). As expected for paracetamol, prolonged GE  
508 translated into absorption delay under fasted conditions (Figure 8 and Figure 9). Under fed conditions,  
509 GE was investigated as a function of a range of caloric contents of an infant meal. Within the current  
510 PBPK modeling exercise and extrapolation from adults to infants, recommended infant formula  
511 volumes and caloric content thereof were used for the estimation of GTT in infants (Eq. 2 and 3) to  
512 facilitate some standardization. PSA performed to understand the uncertainties underlying the caloric  
513 content used in this study demonstrated delayed paracetamol absorption in infants for feeds with  
514 greater caloric contents under infant-formula-fed conditions extrapolated from adults, with less  
515 pronounced sensitivity within the range of 100-200 kcal feed (Figure 8 and Figure 9).

516

517 Paracetamol permeability was another sensitive parameter, the decrease of which led to drug  
518 absorption delay and slight decrease in total exposure (Figures S6, S7, and S8). Interestingly, the PSA  
519 under fed conditions induced with infant formula exhibited less sensitivity towards drug permeability  
520 compared to the fasted conditions regarding  $T_{max}$ . Drug permeability is generally considered to be an  
521 age-dependent factor that reaches maturity the age of 2 years, with most of the conclusions originating  
522 from investigations using dual sugar intestinal permeability tests (3). According to these studies (75–  
523 79), increased permeability has been observed especially during the first days after birth, with  
524 maturation (closure) of the junctions between epithelial cells ranging between the first days after birth  
525 up to 15 months of age. Furthermore, age-dependent changes in permeability could be due to ongoing  
526 morphological development of the shape and size of SI structures, i.e. villi and microvilli, leading to  
527 surface-area-based decreased absorption capacity at young ages (7). While this parameter might bring

528 uncertainty into PBPK models for younger age-groups and should be carefully interpreted, population  
529 pharmacokinetic investigations suggested that age-related changes of paracetamol absorption rate  
530 were prominent in the early days after birth, i.e., neonates, who were not within the target group in  
531 the current investigation (7).

532

533 The present study for paracetamol highlighted the importance of informing the PBPK model during  
534 development with *in vivo* data employing age-relevant formulation and dosing conditions prior to  
535 extrapolation as opposed to using default settings to predict paracetamol oral absorption in infants  
536 (Figure 1). Along with PBPK modeling limitations highlighted and elaborately discussed elsewhere  
537 (3,7,8), specific limitations of the usefulness and applicability of the results from the present  
538 methodology include uncertainty regarding compounds whose bioavailability is affected by bile salt  
539 solubilization, ionizable compounds affected by intraluminal pH changes, drugs with permeability-  
540 limited absorption or transporter-substrates. In the present study, fasted conditions and/or infant-  
541 formula-fed conditions based on the study by Statelova *et al.* resulted in adequate predictions of  
542 paracetamol suspension performance in infants (27,28). In contrast, extrapolation following the  
543 reference meal appeared less useful to predict the observed plasma levels in infants (27,28). Coupling  
544 *in vivo* investigations of age-appropriate dosing conditions in adults with PBPK modeling and  
545 extrapolation to paediatrics provides a practical strategy for paediatric drug formulation testing with  
546 view to the complex interplay of formulation and age-appropriate meal characteristics.

## 547 Concluding remarks

548 Adult clinical data following paracetamol suspension administration under different dosing conditions  
549 was successfully extrapolated to infants using PBPK modeling. Reasonable simulations were achieved  
550 applying the refined model for fasted and/or fed state conditions employing a paracetamol meal-  
551 dependent GE based on infant formula. In contrast, default software parameters (GTT) and  
552 extrapolation to infants using paracetamol GE following the solid-liquid reference meal appeared less  
553 useful for predicting early exposure. The present investigation extended the utilization of PBPK  
554 modeling for simulating plasma concentration levels in infant populations in the context of its  
555 application within the biopharmaceutical investigations of age-appropriate fed conditions. Emphasis  
556 should be placed on age-dependent meal-drug-formulation interactions, as drug formulations for  
557 infants can be different than adults', i.e., suspensions, mini-tablets or multiparticulates and paediatric  
558 meals have commonly homogeneous texture unlike the reference meal. Our findings support the need  
559 of paediatric formulation investigations employing foods commonly used in the target paediatric  
560 subpopulation as recently introduced in regulatory guidelines (11). Furthermore, the present  
561 investigation indicated that caution should be exercised even when using bioavailability data of BCS  
562 Class I drugs with non-problematic absorption in adults to extrapolate to infants. Verification of the  
563 proposed methodology for infant formulation evaluation with broader spectrum of compounds with  
564 different physicochemical properties is required. Finally, availability of high-quality clinical data in  
565 infants is of paramount importance for evaluating the biopharmaceutics tools and methodologies and  
566 confirmation of their reliability.

567

568

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## 574 REFERENCES

- 575 1. United States Congress 107th. Best Pharmaceuticals for children act [Internet]. Vols. 107–109.  
576 2002. p. 1–18. Available from: [https://www.congress.gov/107/plaws/publ109/PLAW-](https://www.congress.gov/107/plaws/publ109/PLAW-107publ109.pdf)  
577 [107publ109.pdf](https://www.congress.gov/107/plaws/publ109/PLAW-107publ109.pdf)
- 578 2. (EU) TEP and TC of the EU. Regulation No 1901/2006. Off J Eur Union [Internet]. 2006;  
579 Available from: [https://ec.europa.eu/health/sites/health/files/files/eudralex/vol-](https://ec.europa.eu/health/sites/health/files/files/eudralex/vol-1/reg_2006_1901/reg_2006_1901_en.pdf)  
580 [1/reg\\_2006\\_1901/reg\\_2006\\_1901\\_en.pdf](https://ec.europa.eu/health/sites/health/files/files/eudralex/vol-1/reg_2006_1901/reg_2006_1901_en.pdf)
- 581 3. Guimarães M, Stelova M, Holm R, Reppas C, Symillides M, Vertzoni M, et al.  
582 Biopharmaceutical considerations in paediatrics with a view to the evaluation of orally  
583 administered drug products - a PEARRL review. *J Pharm Pharmacol*. 2019;71(4):603–42.
- 584 4. Batchelor H, Kaukonen AM, Klein S, Davit B, Ju R, Ternik R, et al. Food effects in paediatric  
585 medicines development for products co-administered with food. *Int J Pharm*. 2018  
586 Feb;536(2):530–5.
- 587 5. Elder DP, Holm R, Kuentz M. Medicines for Pediatric Patients—Biopharmaceutical,  
588 Developmental, and Regulatory Considerations. *J Pharm Sci*. 2017;106(4):950–60.
- 589 6. Kohlmann P, Stillhart C, Kuentz M, Parrott N. Investigating Oral Absorption of Carbamazepine  
590 in Pediatric Populations. *AAPS J*. 2017;19(6):1864–77.
- 591 7. Johnson TN, Bonner JJ, Tucker GT, Turner DB, Jamei M. Development and application of a  
592 physiologically-based model of paediatric oral drug absorption. *Eur J Pharm Sci*. 2018; 115:57-  
593 67
- 594 8. Verscheijden LFM, Koenderink JB, Johnson TN, Wildt SN De, Russel FGM. Physiologically-based  
595 pharmacokinetic models for children: Starting to reach maturation ? *Pharmacol Ther*.  
596 2020;10.
- 597 9. Stelova M, Goumas K, Fotaki N, Holm R, Symillides M, Reppas C, et al. On the Design of Food  
598 Effect Studies in Adults for Extrapolating Oral Drug Absorption Data to Infants : an Exploratory  
599 Study Highlighting the Importance of Infant Food. *AAPS J*. 2020;22(6):1–11.
- 600 10. Maharaj AR, Edginton AN. Physiologically based pharmacokinetic modeling and simulation in  
601 pediatric drug development. *CPT pharmacometrics Syst Pharmacol*. 2014;3(November):e150.
- 602 11. Food and Drug Administration (FDA). Assessing the effects of food on drugs in INDs and NDAs-  
603 clinical pharmacology considerations guidance for industry [Internet]. 2019. Available from:  
604 <http://www.fda.gov/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/default.htm>

- 605 m
- 606 12. Batchelor H. Influence of food on paediatric gastrointestinal drug absorption following oral  
607 administration: a review. *Children*. 2015;2(2):244–71.
- 608 13. European Medicines Agency (EMA). Guideline on the investigation of drug interactions. Guid  
609 Doc [Internet]. 2012;44(June):1–59. Available from:  
610 [http://www.ema.europa.eu/docs/en\\_GB/document\\_library/Scientific\\_guideline/2012/07/WC](http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2012/07/WC500129606.pdf)  
611 [500129606.pdf](http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2012/07/WC500129606.pdf)
- 612 14. Food and Drug Administration (FDA). Guidance for industry food-effect bioavailability and fed  
613 bioequivalence studies. 2002;(December):1–12. Available from:  
614 [https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances](https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm070241.pdf)  
615 [/ucm070241.pdf](https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances/ucm070241.pdf)
- 616 15. Atkinson HC, Stanescu I, Frampton C, Salem II, Beasley CPH, Robson R. Pharmacokinetics and  
617 bioavailability of a fixed-dose combination of ibuprofen and paracetamol after intravenous  
618 and oral administration. *Clin Drug Investig*. 2015;35(10):625–32.
- 619 16. Clements J, Critchley J, Prescott L. The role of sulphate conjugation in the metabolism and  
620 disposition of oral and intravenous paracetamol in man. *Br J Clin Pharmacol*. 1984;18(4):481–  
621 5.
- 622 17. De Morais S, Uetrecht J, Wells P. Decreased glucuronidation and increased bioactivation of  
623 acetaminophen in Gilbert’s syndrome. *Gastroenterology*. 1992;102(2):577–86.
- 624 18. Douglas AP, Savage RL, Rawlins MD. Paracetamol (acetaminophen) kinetics in patients with  
625 Gilbert’s syndrome. *Eur J Clin Pharmacol*. 1978;13(3):209–12.
- 626 19. Eandi M, Viano I, Gamalero SR. Absolute bioavailability of paracetamol after oral or rectal  
627 administration in healthy volunteers. *Drug Res (Stuttg)*. 1984;34(8):903–7.
- 628 20. Liukas A, Kuusniemi K, Aantaa R, Virolainen P, Niemi M, Neuvonen PJ, et al. Pharmacokinetics  
629 of intravenous paracetamol in elderly patients. *Clin Pharmacokinet*. 2011;50(2):121–9.
- 630 21. Perucca E, Richens A. Paracetamol disposition in normal subjects and in patients treated with  
631 antiepileptic drugs. *Br J Clin Pharmacol*. 1979;7(2):201–6.
- 632 22. Prescott LF, Speirs GC, Critchley JA, Temple RM, Winney RJ. Paracetamol disposition and  
633 metabolite kinetics in patients with chronic renal failure. *Eur J Clin Pharmacol*.  
634 1989;36(3):291–7.
- 635 23. Rawlins MD, Henderson DB, Hijab AR. Pharmacokinetics of paracetamol (acetaminophen)

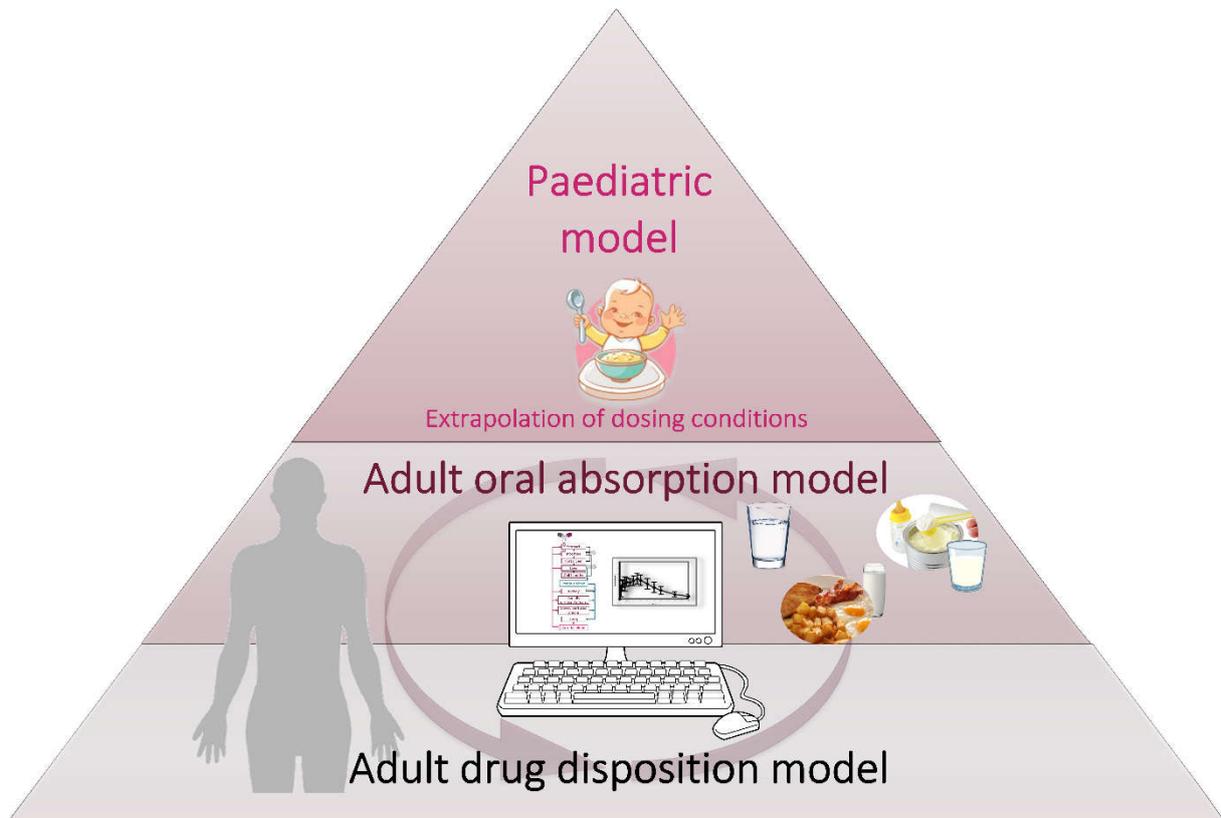
- 636 after intravenous and oral administration. *Eur J Clin Pharmacol.* 1977;11:283–6.
- 637 24. Singla NK, Parulan C, Samson R, Hutchinson J, Bushnell R, Beja EG, et al. Plasma and  
638 Cerebrospinal Fluid Pharmacokinetic Parameters After Single-Dose Administration of  
639 Intravenous, Oral, or Rectal Acetaminophen. *Pain Pract.* 2012;12(7):523–32.
- 640 25. Zuppa A, Hammer G, Barrett J, Kenney B, Kassir N, Moukassi S, et al. Safety and Population  
641 Pharmacokinetic Analysis of Intravenous Acetaminophen in Neonates, Infants, Children, and  
642 Adolescents With Pain or Fever. *J Pediatr Pharmacol Ther.* 2011;16(4):246–61.
- 643 26. Mohammed BS, Engelhardt T, Cameron GA, Cameron L, Hawksworth GM, Hawwa AF, et al.  
644 Population pharmacokinetics of single-dose intravenous paracetamol in children. *Br J*  
645 *Anaesth.* 2012;108(5):823–9.
- 646 27. Hopkins CS, Underhill S, Booker PD. Pharmacokinetics of paracetamol after cardiac surgery.  
647 *Arch Dis Child.* 1990;65(9):971–6.
- 648 28. Walson PD, Halvorsen M, Edge J, Casavant MJ, Kelley MT. Pharmacokinetic Comparison of  
649 Acetaminophen Elixir Versus Suppositories in Vaccinated Infants (Aged 3 to 36 Months): A  
650 Single-Dose, Open-Label, Randomized, Parallel-Group Design. *Clin Ther.* 2013;35(2):135–40.
- 651 29. Tsamandouras N, Rostami-Hodjegan A, Aarons L. Combining the “bottom up” and “top down”  
652 approaches in pharmacokinetic modelling: Fitting PBPK models to observed clinical data. *Br J*  
653 *Clin Pharmacol.* 2015;79(1):48–55.
- 654 30. Jamei M. Recent Advances in Development and Application of Physiologically-Based  
655 Pharmacokinetic (PBPK) Models: a Transition from Academic Curiosity to Regulatory  
656 Acceptance. *Curr Pharmacol Reports.* 2016;2(3):161–9.
- 657 31. Kalantzi L, Reppas C, Dressman JB, Amidon GL, Junginger HE, Midha KK, et al. Biowaiver  
658 monographs for immediate release solid oral dosage forms: Acetaminophen (Paracetamol). *J*  
659 *Pharm Sci.* 2006;95(1):4–14.
- 660 32. Jiang X-L, Zhao P, Barrett JS, Lesko LJ, Schmidt S. Application of physiologically based  
661 pharmacokinetic modeling to predict acetaminophen metabolism and pharmacokinetics in  
662 children. *CPT pharmacometrics Syst Pharmacol.* 2013;2(August):e80.
- 663 33. Ladumor MK, Bhatt DK, Gaedigk A, Sharma S, Thakur A, Pearce RE, et al. Ontogeny of hepatic  
664 sulfotransferases and prediction of age-dependent fractional contribution of sulfation in  
665 acetaminophen metabolism. *Drug Metab Dispos.* 2019;47(8):818–31.
- 666 34. Yamashita S, Furubayashi T, Kataoka M, Sakane T, Sezaki H, Tokuda H. Optimized conditions  
667 for prediction of intestinal drug permeability using Caco-2 cells. *Eur J Pharm Sci.*

- 668 2000;10(3):195–204.
- 669 35. Sun D, Lennernas H, Welage LS, Barnett JL, Landowski CP, Foster D, et al. Comparison of  
670 human duodenum and Caco-2 gene expression profiles for 12,000 gene sequences tags and  
671 correlation with permeability of 26 drugs. *Pharm Res.* 2002;19(10):1400–16.
- 672 36. Strougo A, Eissing T, Yassen A, Willmann S, Danhof M, Freijer J. First dose in children:  
673 Physiological insights into pharmacokinetic scaling approaches and their implications in  
674 paediatric drug development. *J Pharmacokinet Pharmacodyn.* 2012;39(2):195–203.
- 675 37. Prescott L. Kinetics and metabolism of paracetamol and phenacetin. *Br J Clin Pharmacol.*  
676 1980;10(2 S):291S-298S.
- 677 38. Rodgers T, Rowland M. Physiologically Based Pharmacokinetic Modelling 2: Predicting the  
678 Tissue Distribution of Acids, Very Weak Bases, Neutrals and Zwitterions. *J Pharm Sci.*  
679 2005;95(6):1238–57.
- 680 39. Lukacova V, Parrott NJ, Lave T, Fraczekiewicz G, Bolger MB. Role of fraction unbound in plasma  
681 in calculation of tissue:plasma partition coefficients. In: AAPS National meeting, Atlanta,  
682 November 15-20. 2008.
- 683 40. Laine JE, Auriola S, Pasanen M, Juvonen RO. Acetaminophen bioactivation by human  
684 cytochrome P450 enzymes and animal microsomes. *Xenobiotica.* 2009;39(1):11–21.
- 685 41. Mutlib AE, Goosen TC, Bauman JN, Williams JA, Kulkarni S, Kostrubsky S. Kinetics of  
686 acetaminophen glucuronidation by UDP-glucuronosyltransferases 1A1, 1A6, 1A9 and 2B15.  
687 Potential implications in acetaminophen-induced hepatotoxicity. *Chem Res Toxicol.*  
688 2006;19(5):701–9.
- 689 42. Adjei AA, Gaedigk A, Simon SD, Weinshilboum RM, Leeder JS. Interindividual variability in  
690 acetaminophen sulfation by human fetal liver: Implications for pharmacogenetic  
691 investigations of drug-induced birth defects. *Birth Defects Res Part A - Clin Mol Teratol.*  
692 2008;82(3):155–65.
- 693 43. SimulationsPlus Inc. GastroPlus Manual.
- 694 44. T’jollyn H, Vermeulen A, Van Bocxlaer J. PBPK and its Virtual Populations: the Impact of  
695 Physiology on Pediatric Pharmacokinetic Predictions of Tramadol. *AAPS J.* 2019;21(1):1–12.
- 696 45. Villiger A, Stillhart C, Parrott N, Kuentz M. Using Physiologically Based Pharmacokinetic (PBPK)  
697 Modelling to Gain Insights into the Effect of Physiological Factors on Oral Absorption in  
698 Paediatric Populations. *AAPS J.* 2016;18(4):933–47.

- 699 46. Edginton AN, Willmann S. Physiology-based Versus Allometric Scaling Of Clearance In  
700 Children : A Comparison. 2006;51368.
- 701 47. Lowenthal DT, Oie S, van Stone JC, Briggs WA, Levy G. Pharmacokinetics of acetaminophen  
702 elimination by anephric patients. J Pharmacol Exp Ther. 1976;196(3):570–8.
- 703 48. Terry SI, Gould JC, McManus JPA, Prescott LF. Absorption of penicillin and paracetamol after  
704 small intestinal bypass surgery. Eur J Clin Pharmacol. 1982;23(3):245–8.
- 705 49. Boase S, Miners JO. In vitro-in vivo correlations for drugs eliminated by glucuronidation:  
706 Investigations with the model substrate zidovudine. Br J Clin Pharmacol. 2002;54(5):493–503.
- 707 50. Agoram B, Woltosz WS, Bolger MB. Predicting the impact of physiological and biochemical  
708 processes on oral drug bioavailability. Adv Drug Deliv Rev. 2001;50:S41–67.
- 709 51. Kostewicz ES, Aarons L, Bergstrand M, Bolger MB, Galetin A, Hatley O, et al. PBPK models for  
710 the prediction of in vivo performance of oral dosage forms. Eur J Pharm Sci. 2014;57(1):300–  
711 21.
- 712 52. Willems M, Quartero AO, Numans ME. How useful is paracetamol absorption as a marker of  
713 gastric emptying? A systematic literature study. Dig Dis Sci. 2001;46(10):2256–62.
- 714 53. Johnson TN, Rostami-hodjegan A, Tucker GT. Prediction of the Clearance of Eleven Drugs and  
715 Associated Variability in Neonates , Infants and Children. 2006;45(9):931–56.
- 716 54. Badée J, Qiu N, Collier AC, Takahashi RH, Forrest WF, Parrott N, et al. Characterization of the  
717 Ontogeny of Hepatic UDP-Glucuronosyltransferase Enzymes Based on Glucuronidation  
718 Activity Measured in Human Liver Microsomes. J Clin Pharmacol. 2019 Sep;59(S1):S42–55.
- 719 55. Bonner JJ, Vajjah P, Abduljalil K, Jamei M, Rostami-Hodjegan A, Tucker GT, et al. Does age  
720 affect gastric emptying time? A model-based meta-analysis of data from premature neonates  
721 through to adults. Biopharm Drug Dispos. 2015;36(4):245–57.
- 722 56. Obach RS, Baxter JG, Liston TE, Silber BM, Jones BC, Macintyre F, et al. The Prediction of  
723 Human Pharmacokinetic Parameters from Preclinical and In Vitro Metabolism Data. J  
724 Pharmacol Exp Ther. 1997 Oct 1;283(1):46 LP – 58.
- 725 57. Mahmood I, Ahmad T, Mansoor N, Sharib SM. Prediction of Clearance in Neonates and Infants  
726 ( $\leq 3$  Months of Age) for Drugs That Are Glucuronidated: A Comparative Study Between  
727 Allometric Scaling and Physiologically Based Pharmacokinetic Modeling. J Clin Pharmacol.  
728 2017;57(4):476–83.
- 729 58. Lennernäs H. Human *in Vivo* Regional Intestinal Permeability: Importance for Pharmaceutical

- 730 Drug Development. *Mol Pharm.* 2014;11(1):12–23.
- 731 59. Levitt DG. Quantitation of small intestinal permeability during normal human drug absorption.  
732 *BMC Pharmacol Toxicol.* 2013;14(1):1.
- 733 60. Sanaka M, Kuyama Y, Shimomura Y, Saitoh M, Hattori K. New Mathematical Model for  
734 Accurate Description of Absorption Kinetics of Paracetamol Given Orally With a High Calorie  
735 Liquid Meal. *Int J Clin Pharmacol Ther.* 2002;40(11):499–506.
- 736 61. Bermejo M, Hens B, Dickens J, Mudie D, Paix P. A Mechanistic Physiologically-Based  
737 Biopharmaceutics Modeling ( PBBM ) Approach to Assess the In Vivo Performance of an Orally  
738 Administered Drug Product : From IVIVC to IVIVP. *Pharmaceutics.* 2020;12(1):74.
- 739 62. Van Den Abeele J, Brouwers J, Tack J, Augustijns P. Exploring the link between gastric motility  
740 and intragastric drug distribution in man. *Eur J Pharm Biopharm.* 2017;112:75–84.
- 741 63. Grimm M, Scholz E, Koziolk M, Kuhn JP, Weitschies W. Gastric water emptying under fed  
742 state clinical trial conditions is as fast as under fasted conditions Gastric water emptying  
743 under fed state clinical trial conditions is as fast as under fasted conditions. *Mol Pharm.*  
744 2017;14(12):4262–71.
- 745 64. Pentafragka C, Vertzoni M, Symillides M, Goumas K, Reppas C. Disposition of two highly  
746 permeable drugs in the upper gastrointestinal lumen of healthy adults after a standard high-  
747 calorie, high-fat meal. *Eur J Pharm Sci.* 2020;149(January).
- 748 65. Van Den Abeele J, Rayyan M, Hoffman I, Van de Vijver E, Zhu W, Augustijns P. Gastric fluid  
749 composition in a paediatric population: Age-dependent changes relevant for gastrointestinal  
750 drug disposition. *Eur J Pharm Sci.* 2018;123:301–11.
- 751 66. Yeung CHT, Fong S, Malik PRV, Edginton AN. Quantifying breast milk intake by term and  
752 preterm infants for input into paediatric physiologically based pharmacokinetic models.  
753 *Matern Child Nutr.* 2020;16(2):1–33.
- 754 67. Koziolk M, Alcaro S, Augustijns P, Basit AW, Grimm M, Hens B, et al. The mechanisms of  
755 pharmacokinetic food-drug interactions – A perspective from the UNGAP group. *Eur J Pharm*  
756 *Sci.* 2019;134(March):31–59.
- 757 68. Hauser B, Roelants M, De Schepper J, Veereman G, Caveliers V, Devreker T, et al. Gastric  
758 Emptying of Liquids in Children. *J Pediatr Gastroenterol Nutr.* 2015;62(3):403–8.
- 759 69. Lange A, Funch-Jensen P, Thommesen P, Schiøtz PO. Gastric emptying patterns of a liquid  
760 meal in newborn infants measured by epigastric impedance. *Neurogastroenterol Motil.*  
761 1997;9(2):55–62.

- 762 70. Johnson TN, Bonner JJ, Tucker GT, Turner DB, Jamei M. Development and applications of a  
763 physiologically-based model of paediatric oral drug absorption. *Eur J Pharm Sci.*  
764 2018;115(June 2017):57–67.
- 765 71. Billeaud C, Guillet J, Sandler B. Gastric emptying in infants with or without gastroesophageal  
766 reflux according to the type of milk. *Eur J Clin Nutr.* 1990;44(September 1990):577–83.
- 767 72. Cavell B. Gastric emptying in infants with cystic fibrosis. *Acta Paediatr Scand.* 1981;70:635–8.
- 768 73. Staelens S, Van Den Driessche M, Barclay D, Carrié-Faessler AL, Haschke F, Verbeke K, et al.  
769 Gastric emptying in healthy newborns fed an intact protein formula, a partially and an  
770 extensively hydrolysed formula. *Clin Nutr.* 2008;27(2):264–8.
- 771 74. Van Den Driessche M, Veereman-Wauters G. Gastric emptying in infants and children. *Acta*  
772 *Gastroenterol Belg.* 2003;66(4):274–82.
- 773 75. Lee GO, McCormick BJJ, Seidman JC, Kosek MN, Haque R, Olortegui MP, et al. Infant  
774 Nutritional Status, Feeding Practices, Enteropathogen Exposure, Socioeconomic Status, and  
775 Illness Are Associated with Gut Barrier Function As Assessed by the Lactulose Mannitol Test in  
776 the MAL-ED Birth Cohort. *Am J Trop Med Hyg.* 2017 Jul;97(1):281–90.
- 777 76. Riezzo G, Indrio F, Raimondi F, Montagna O, Salvia G, Massimo B, et al. Maturation of gastric  
778 electrical activity, gastric emptying and intestinal permeability in preterm newborns during  
779 the first month of life. *Ital J Pediatr.* 2009 Mar 15;35(1):6.
- 780 77. Corpeleijn WE, van Vliet I, de Gast-Bakker D-AH, van der Schoor SRD, Alles MS, Hoijer M, et al.  
781 Effect of Enteral IGF-1 Supplementation on Feeding Tolerance, Growth, and Gut Permeability  
782 in Enterally Fed Premature Neonates. *J Pediatr Gastroenterol Nutr.* 2008;46(2).
- 783 78. Saleem B, Okogbule-Wonodi AC, Fasano A, Magder LS, Ravel J, Kapoor S, et al. Intestinal  
784 Barrier Maturation in Very Low Birthweight Infants: Relationship to Feeding and Antibiotic  
785 Exposure. *J Pediatr.* 2017;183:31-36.e1.
- 786 79. Shulman RJ, Schanler RJ, Lau C, Heitkemper M, Ou C-N, Smith EO. Early Feeding, Antenatal  
787 Glucocorticoids, and Human Milk Decrease Intestinal Permeability in Preterm Infants. *Pediatr*  
788 *Res.* 1998;44(4):519–23.
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## List of figures

**Figure 1** Model development strategy for the evaluation of food effects in infants based on *in vivo* data in adults. Adapted from (3).

**Figure 2** Simulations of paracetamol plasma concentrations following i.v. administration in healthy adults. The disposition model was developed according to data observed at a low (A) (5 mg/kg, i.e. 350 mg) and high dose (B) (20 mg/kg, i.e. 1400 mg) (16). Model verification was performed with clinical data sets not used during model development at low 500 mg (C) and high 1000 mg (D) doses (15). Symbols denote observed mean data, error bars represent the standard deviation of the observed data, and continuous lines represent the simulated plasma concentration-time profile.

**Figure 3** Simulations of paracetamol plasma concentrations following oral administration of paracetamol drops solution (A) and solution (B) to healthy adults at a dose of 1000 mg according to (19,21). Symbols denote observed mean data, error bars represent the standard deviation of the observed data, and discontinuous lines represent the simulated plasma concentration-time profile.

**Figure 4** Predicted plasma concentration-time profiles (continuous purple line) following oral administration of pediatric suspension under different dosing conditions: fasted conditions employing default GTT value 0.1 h (A) and adjusted GTT value of 0.75 h according to *in vivo* observations (B); Reference-meal-fed conditions employing default calorie-based software estimated GTT of 3.43 h (C) and adjusted GTT of 1.5 h according to *in vivo* observations (D) with first order GE; and infant-formula-fed conditions simulating infant dosing employing default calorie-based software estimated GTT 2.03 (E) and adjusted GTT of 4.5 h (F) with zero-order GE. Grey lines denote individual observed data and symbols and error bars denote mean observed plasma levels and the standard deviation (n=8 healthy male adult volunteers) (9).

**Figure 5** Simulated plasma concentration-time profiles (continuous purple lines) in infants (A and B) and in children (C and D) after i.v. administration of paracetamol at doses 15 mg/kg (A and C) or 12.5 mg/kg (B and D). Observed mean concentrations and standard deviations depicted as black symbols and error bars, individual concentrations (n=25 infants and n=56 children and adolescents) are depicted with open symbols (25).

**Figure 6** Predicted plasma concentration-time profiles (purple lines) in infants under software default fasted conditions, i.e. GTT 0.1 h (A) and adjusted fasted conditions, i.e. GTT 0.75 h (B); fed conditions employing first order GE (solid-liquid meal) and software default GTT value of 2.1 h (C) or adjusted GTT value of 0.21 h (D); infant-formula-fed conditions following zero order GE kinetics (liquid homogeneous meals) using software default GTT value of 2.1 h (E) or adjusted GTT value of 1.21 (F). Observed mean concentrations and standard deviations depicted as symbols and error bars, individual observed data is presented with grey lines (27).

**Figure 7** Predicted plasma concentration-time profiles (purple lines) in infants under software default fasted conditions, i.e. GTT 0.1 h (A) and adjusted fasted conditions, i.e. GTT 0.75 h (B); fed conditions employing first order GE (solid-liquid meal) employing software default GTT value of 1.89 h (C) and adjusted GTT value of 0.26 h (D); infant-formula-fed conditions following zero order GE kinetics (liquid homogeneous meals) using software default GTT value of 1.89 h (E) and adjusted GTT value of 1.47 (F). Observed mean concentrations depicted as symbols (28).

**Figure 8** Simulated plasma concentration-time profile (continuous line) in infant population representative under fasted conditions with variation of the GTT between 0.1-1.5 h (A), under fed conditions with different caloric intake (70-200 kcal) and adjusted GTT based on the paracetamol reference-meal dependent gastric emptying (B), and under infant fed conditions with different caloric intake (70-200 kcal) and adjusted GTT extrapolated based on the paracetamol infant-formula-dependent gastric emptying (C). The color gradient represents increasing GTT values and caloric content of the meals from dark to light grey. Observed mean data and standard deviation are presented as symbols and error bars (27).

**Figure 9** Parameter sensitivity analysis for C<sub>max</sub> and T<sub>max</sub> results in a population representative infant (4 months old, (27)) as a function of GTT under adjusted fasted conditions (A and C), or caloric content (GTT) influence under adjusted reference-meal-conditions (continuous lines B and D), or caloric content influence under adjusted infant-formula-fed conditions (discontinuous lines B and D). Values used within the paediatric simulations employing adjusted MGTT values are shown as open circles.

Figure 1

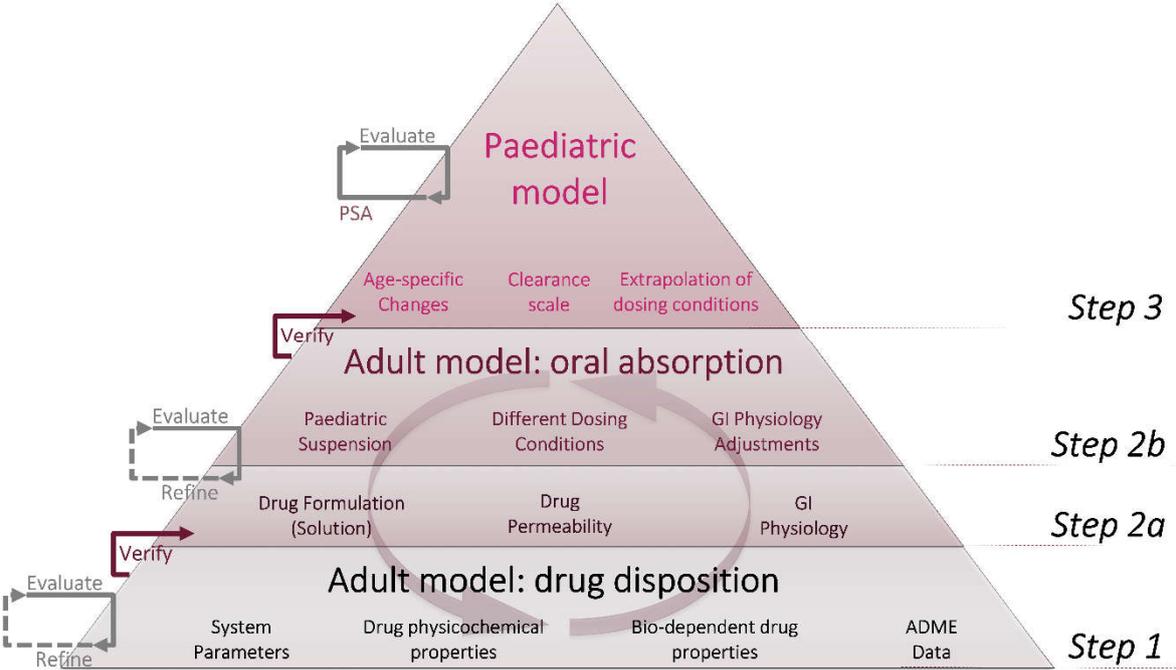


Figure 2

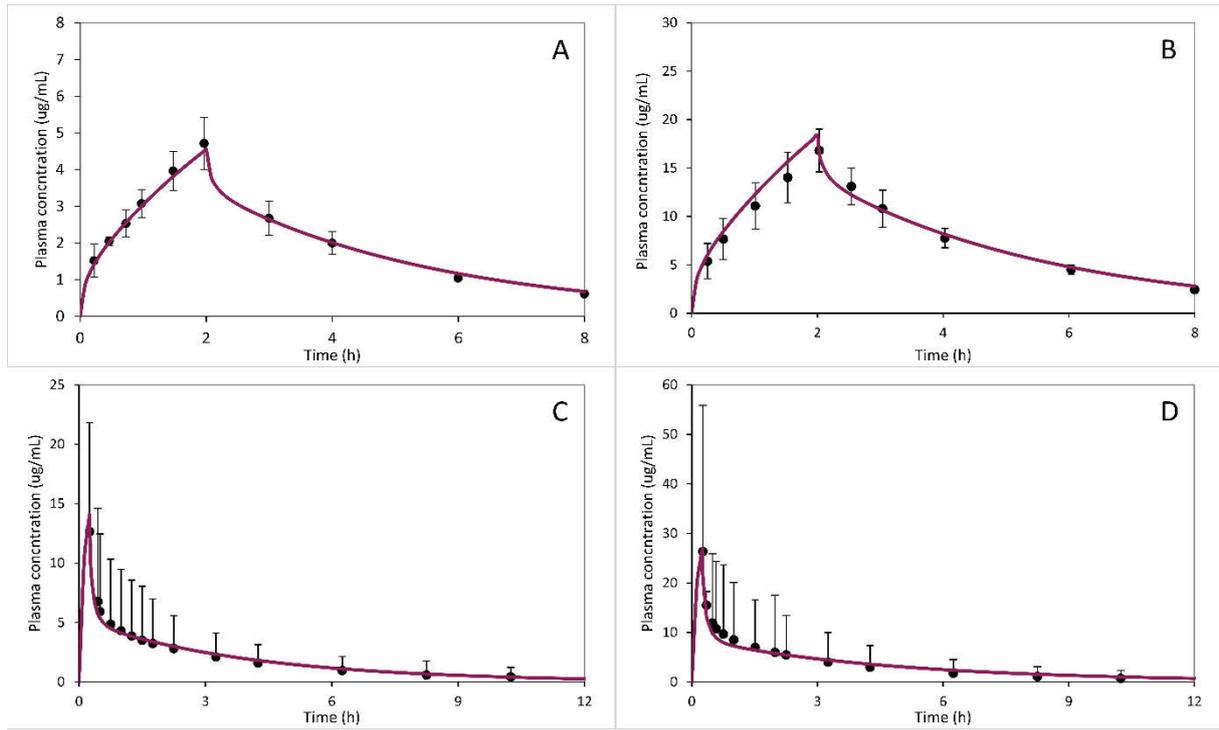


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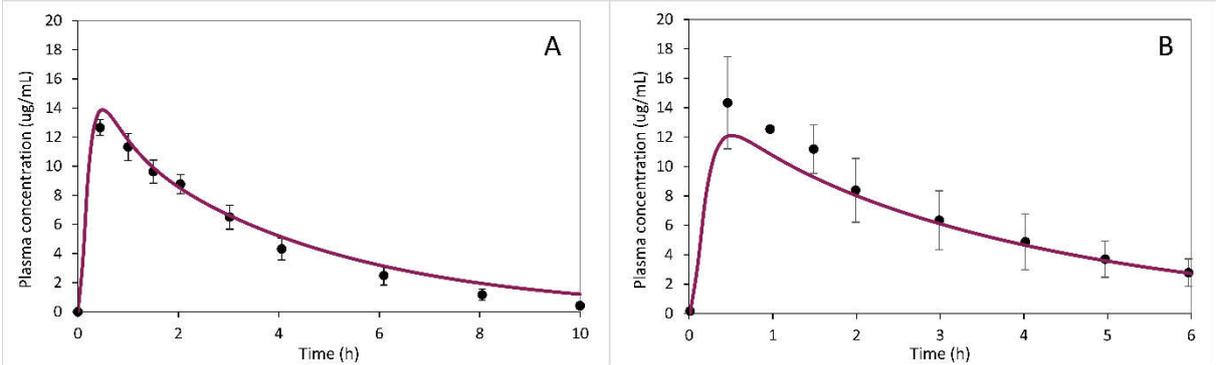


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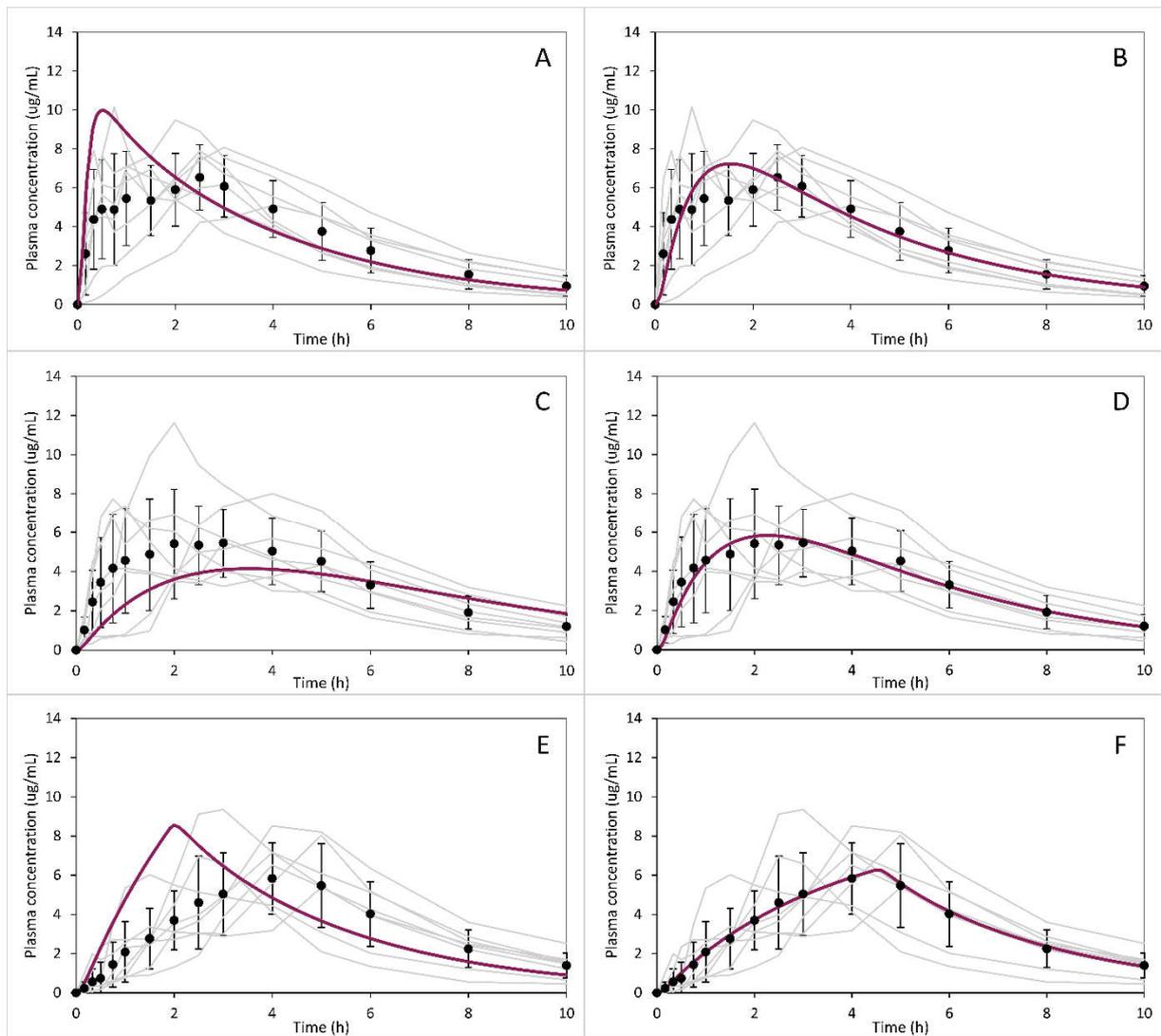


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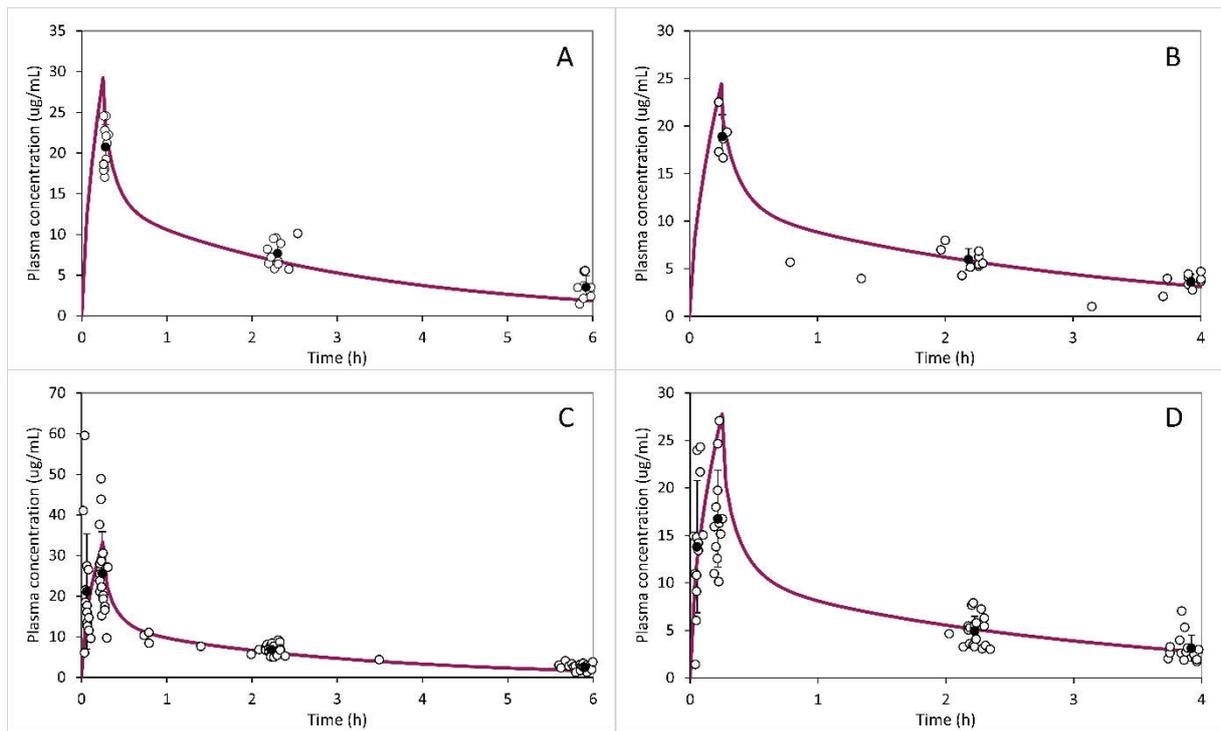


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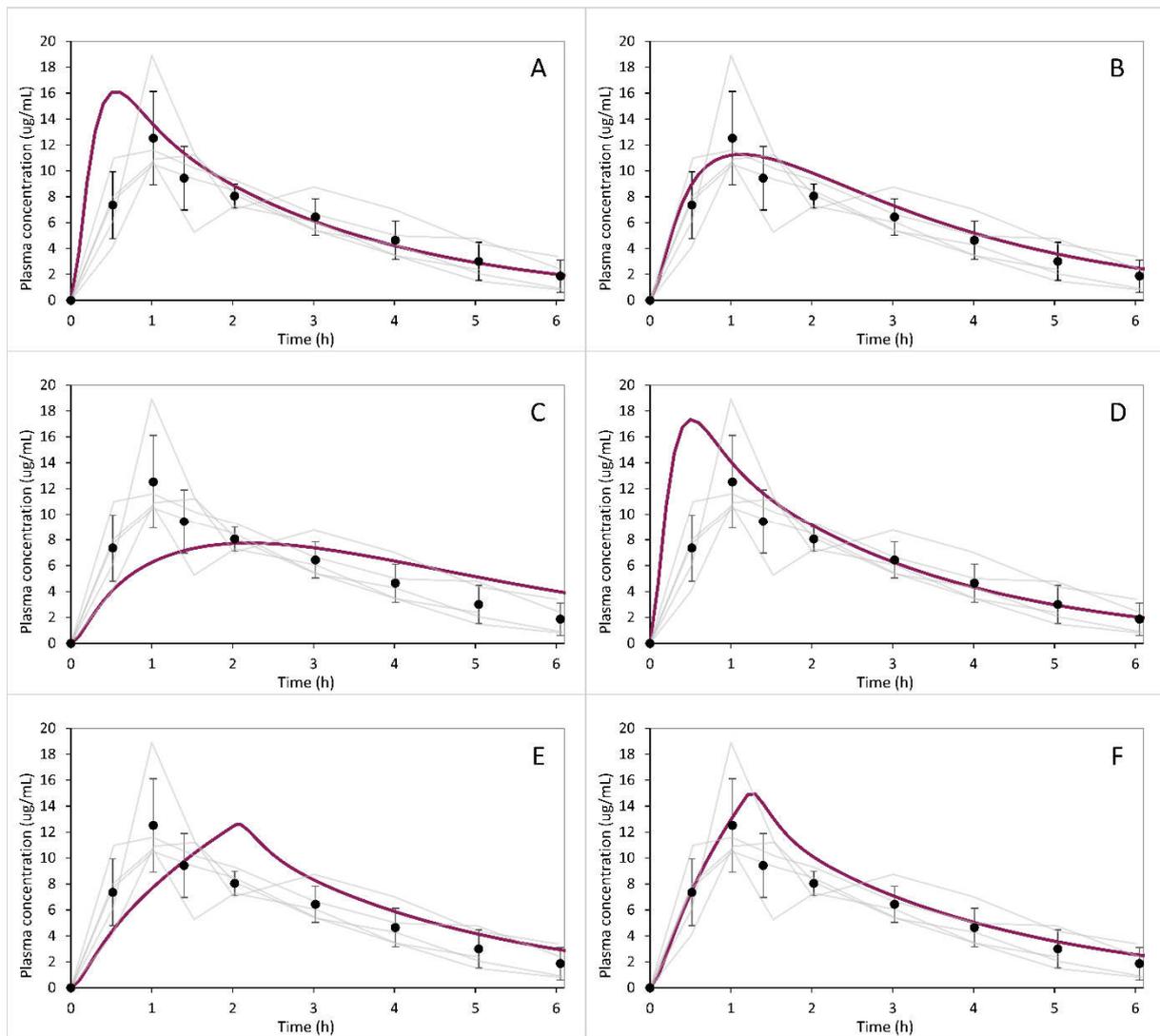


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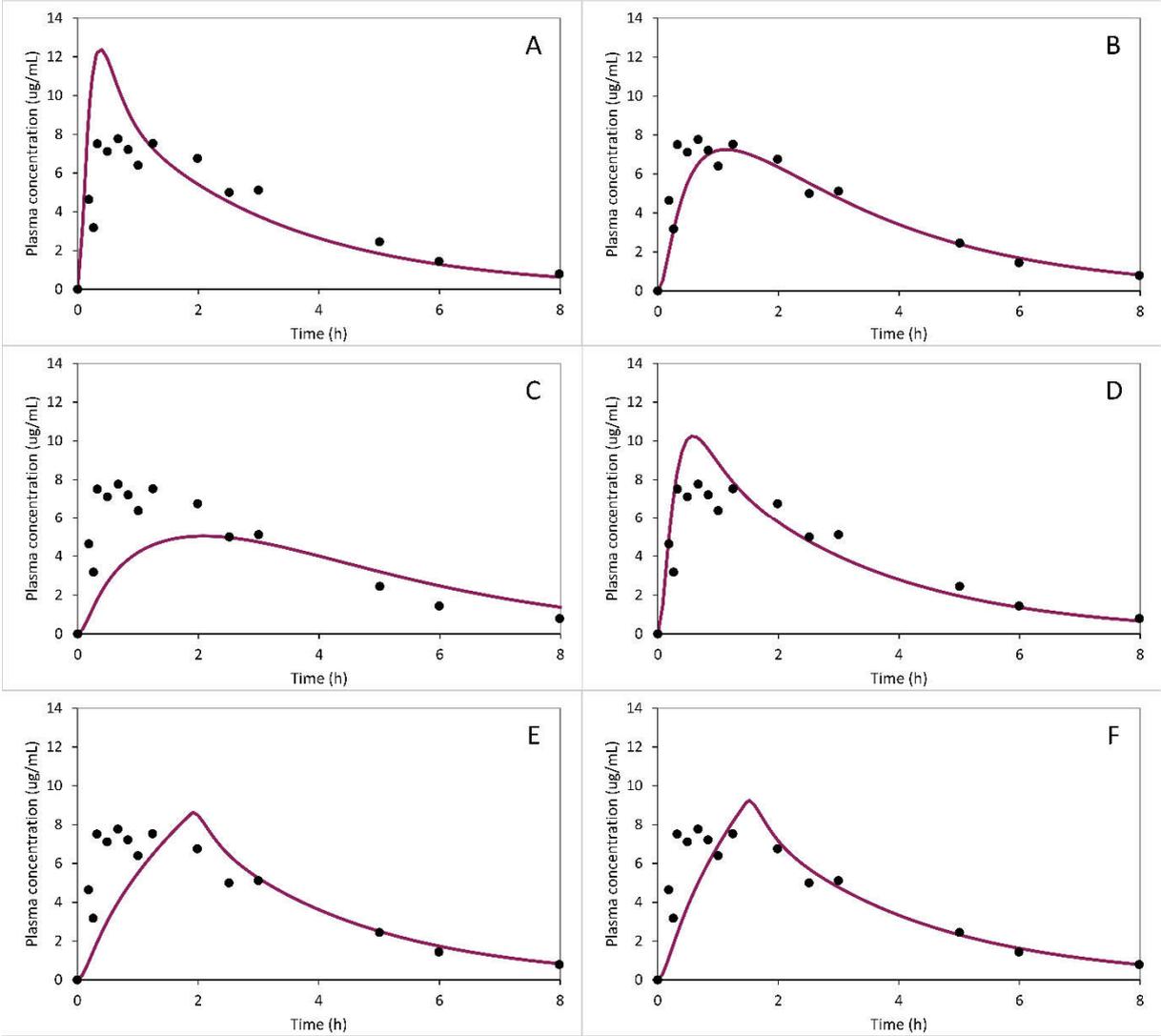


Figure 8

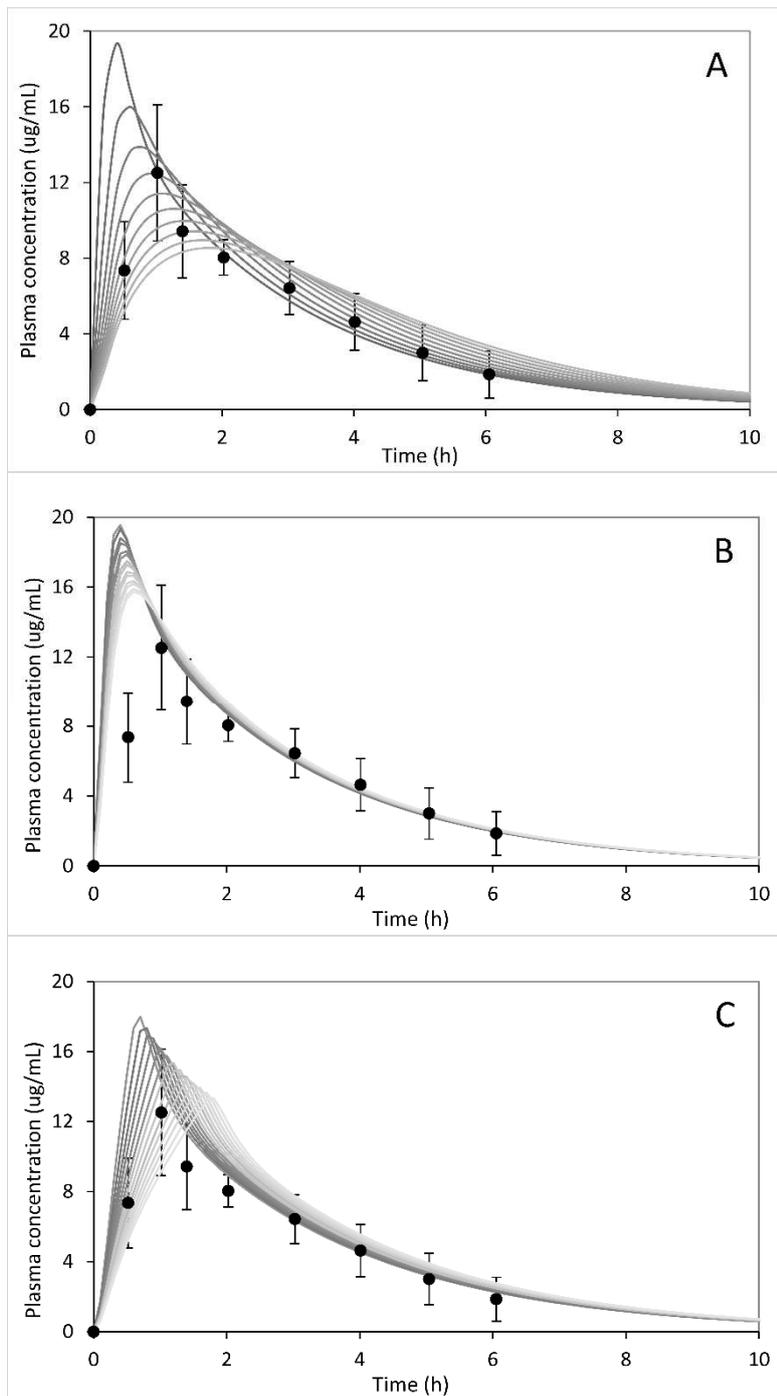
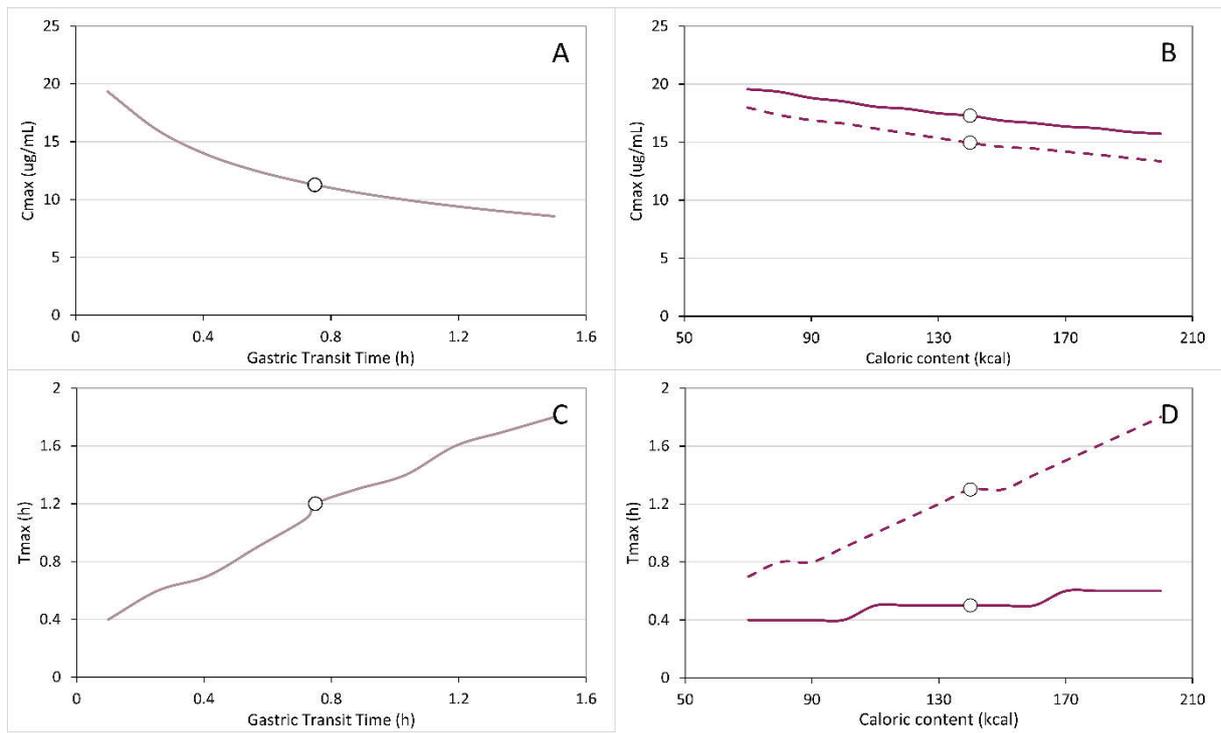


Figure 9



**Table I** Input parameters used to build the PBPK model for paracetamol

Parameter		Source	
<b>Physicochemical properties</b>			
Molecular weight (g/mol)	151.9	(31–33)	
Compound type	Monoprotic weak acid	(31–33)	
pKa	9.45 (acidic)	(31–33)	
logP <sup>a</sup>	0.51	(31–33)	
Reference solubility in water (mg/mL)	14	(31)	
<b>Absorption</b>			
Model	ACAT	GastroPlus™	
Effective permeability, human (cm/s ×10 <sup>4</sup> )	3.897	Calculated based on (7,34,35)	
Dissolution model	Johnson	GastroPlus™	
Drug particle radius (µm)	25	Default GastroPlus™	
<b>Distribution</b>			
Fraction unbound, fu	0.82	(46)	
Blood-plasma ratio	1.09	(47)	
Predicted Vss (L/kg) <sup>b</sup>	0.86	Predicted using the Lukacova, Rodgers and Rowland method (6,38,39)	
<b>Clearance</b>			
<i>In vivo</i> clearance (L/h)	19.7	(16)	
<b>Enzyme kinetics</b>			
	Km (µM)	Vmax (pmol/min/mg microsomal protein)	
CYP1A2 <sup>c</sup>	220	30.78	(40)
CYP2C9 <sup>c</sup>	660	8.42	(40)
CYP2C19 <sup>c</sup>	2000	25.53	(40)
CYP2D6 <sup>c</sup>	440	5.62	(40)
CYP2E1 <sup>c</sup>	4020	76.97	(40)
CYP3A4 <sup>c</sup>	130	57.16	(40)
UGT1A1 <sup>d</sup>	5500	6102.67	(41)
UGT1A9 <sup>d</sup>	9200	10208.11	(41)
UGT2B15 <sup>d</sup>	23000	34045.84	(41)
SULT1A1 <sup>e</sup>	2400	1374.06	(42)
SULT1A3 <sup>e</sup>	1500	202.89	(42)
SULT1E1 <sup>e</sup>	1900	146.22	(42)
SULT2A1 <sup>e</sup>	3700	828.35	(42)

<sup>a</sup> to achieve the benchmark Vss values observed *in vivo*, initially logP value of 1.2 was used for the calculation of the tissue partitioning coefficients (Kp) (36); measured logP value 0.51 was used thought simulations; <sup>b</sup> Predicted volume of distribution at steady state (Vss); <sup>c</sup> Cytochrome P450 (CYP) isoenzyme, <sup>d</sup> UDP-glucuronosyltransferase (UGT) isoenzyme, and <sup>e</sup> cytosolic sulfotransferases (SULT) isoenzyme contributing to paracetamol metabolism

**Table II** Paracetamol meal-dependent gastric emptying (GE) based on the gastric transit time (GTT) values employed in the refined adult model for the reference meal and the infant formula used for inducing fed and infant-formula-fed conditions (9) and adjusted GTT values for paracetamol gastric emptying in infants according to recommended meal calories for age (4 and 10 months).

Meal and Paracetamol GE kinetics	Adult		Paracetamol GE (meal-dependent, expressed as kcal/min)	Infants			
	28-years-old male, 78 kg body weight <sup>a</sup>			4-month-old, 4 kg body weight <sup>b</sup>		10-month-old, 8.6 kg body weight <sup>c</sup>	
	Caloric content (kcal)	GTT (h)		Caloric content (kcal)	GTT (h)	Caloric content (kcal)	GTT (h)
Reference meal (Solid-liquid) 1 <sup>st</sup> order GE	990	1.5	11	140	0.21	170	0.26
Infant formula (Liquid homogeneous) Zero order GE	520	4.5	1.93	140	1.21	170	1.47

<sup>a</sup> mean adult population representative of the study by Statelova *et al.* (9)

<sup>b</sup> mean infant population representative of the study by Hopkins *et al.* (27)

<sup>c</sup> mean infant population representative of the study by Walson *et al.* (28)

**Table III** Observed and predicted pharmacokinetic parameters in studies performed in infants (27,28). Simulations in infants were extrapolated based on the refined adult model for different dosing conditions as described in Stelova *et al.* (9).

Study	Parameter	Observed	Simulated fasted conditions <sup>a</sup>			Simulated fed conditions (solid-liquid meal) <sup>a</sup>			Simulated infant-formula-fed condition (liquid homogenous meal) <sup>a</sup>		
			Predicted	$FD_{pred/obs}$ <sup>b</sup>	$AFE^c / AAFE^d$	Predicted	$FD_{pred/obs}$ <sup>b</sup>	$AFE^c / AAFE^d$	Predicted	$FD_{pred/obs}$ <sup>b</sup>	$AFE^c / AAFE^d$
Hopkins <i>et al.</i> n= 5 subjects 3 male/2 female Dose 19.6 mg/kg	AUC <sub>0-t</sub> <sup>e</sup> (ug/mL·h)	35.93	40.49	1.127	1.129/ 1.187	43.78	1.219	1.053/ 1.201	41.30	1.149	1.212/ 1.215
	AUC <sub>0-inf</sub> <sup>f</sup> (ug/mL·h)	40.21	47.22	1.172		49.24	1.225		49.06	1.220	
	C <sub>max</sub> <sup>g</sup> (ug/mL)	12.52	11.27	0.900		17.33	1.384		14.94	1.193	
	T <sub>max</sub> <sup>h</sup> (h)	1.0	1.1	1.1		0.5	0.5		1.3	1.3	
	AUC <sub>0-T<sub>max</sub></sub> <sup>i</sup> (ug/mL·h)	6.88	7.48	1.087		13.51	1.963		6.83	0.992	
Walson <i>et al.</i> n= 13 subjects 7 male/5 female Dose 12.1 mg/kg	AUC <sub>0-t</sub> <sup>e</sup> (ug/mL·h)	30.13	28.60	0.949	0.948/ 1.201	29.15	0.967	1.107/ 1.274	30.92	1.026	0.847/ 1.401
	AUC <sub>0-inf</sub> <sup>f</sup> (ug/mL·h)	32.76	30.89	0.943		30.99	0.946		28.72	0.876	
	C <sub>max</sub> <sup>g</sup> (ug/mL)	7.77	7.26	0.934		10.24	1.318		9.25	1.190	
	T <sub>max</sub> <sup>h</sup> (h)	0.70	1.12	1.60		0.56	0.80		1.52	2.17	
	AUC <sub>0-T<sub>max</sub></sub> <sup>i</sup> (ug/mL·h)	3.62	2.65	0.733		5.05	1.396		1.78	0.492	

<sup>a</sup> Conditions simulated based on the refined adult model for different dosing condition as described in Stelova *et al.* (9)

<sup>b</sup>  $FD_{pred/obs}$ : Fold difference predicted/observed

<sup>c</sup>  $AFE$  average fold error

<sup>d</sup>  $AAFE$  absolute average fold error

<sup>e</sup> Area under the plasma concentration-time curve from 0h until the last observed time point (t) AUC<sub>0-t</sub> (ug/mL·h)

<sup>f</sup> Area under the plasma concentration-time curve from 0h to infinity AUC<sub>0-inf</sub> (ug/mL·h)

<sup>g</sup> Maximum plasma concentration C<sub>max</sub> (ug/mL)

<sup>h</sup> Time to reach C<sub>max</sub> (h)

<sup>i</sup> Area under the plasma concentration-time curve from 0h until the mean T<sub>max</sub> observed in the simulated study AUC<sub>0-T<sub>max</sub></sub> (ug/mL·h)

